



Microbial Perspectives on Polythene Biodegradation: Exploring the Role of Microorganisms in Addressing Plastic Pollution

**M. Muthukumar^{a*}, A. Aswartha Narayana^b,
A. Dilip Babu^{c++}, Amogha K.R^d, Wankasaki Lytand^{e#},
G. Gomadhi^{ft}, S. Jaya Prabhavathi^{g‡}, G. Malathi^{h^}
and Abhijit Debnath^{i##}**

^a Vivekananda College (Autonomous), (Affiliated to the University of Madras), Chennai - 600004, Tamil Nadu, India.

^b Faculty of Aquaculture, Andhra Kesari University Ongole, Andhra Pradesh -523001, India.

^c Department of Zoology and Aquaculture, Acharya Nagarjuna University Guntur, Andhra Pradesh 522510, India.

^d Department of Aquaculture, College of Fisheries, Mangalore, Karnataka Veterinary Animal and Fisheries Science University, Bidar, India.

^e Department of Microbiology, Shillong College, Shillong, India.

^f Krishi Vigyan Kendra, Tindivanam, Villupuram district, Tamil Nadu, Pin Code: 604 102, India.

^g Regional Research Station, Tamil Nadu Agricultural University, Vridhachalam, 606 001 Cuddalore District, Tamil Nadu, India.

^h Krishi Vigyan Kendra, Sandhiyur, Salem - 636 203, Tamil Nadu, India.

ⁱ Krishi Vigyan Kendra Dhalai, Tripura, -799278 India.

Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/MRJI/2024/v34i51443

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/113576>

⁺⁺ Research Scholar;

[#] Assistant Professor;

[†] Associate Professor (Soil Science and Agrl. Chemistry);

[‡] Associate Professor (Agricultural Entomology);

[^] Associate Professor (Horticulture);

^{##} Subject Matter Specialist;

*Corresponding author: E-mail: mmuthukumar@rkmvc.ac.in;

ABSTRACT

Plastic pollution, particularly from polythene (polyethylene), has emerged as a significant environmental concern worldwide. In response to this challenge, microbial perspectives on polythene biodegradation have garnered attention as potential solutions to mitigate plastic pollution. This article provides an overview of the mechanisms underlying microbial polythene biodegradation, including surface erosion, biofilm formation, metabolic pathways, synergistic interactions, and adaptation. Furthermore, it explores the diversity of polythene-degrading microorganisms and their roles in plastic degradation across different environments. Environmental factors influencing polythene biodegradation, such as temperature, pH, moisture, and nutrient availability, are discussed, along with strategies to optimize degradation rates. Biotechnological approaches, including microbial consortia development and genetic engineering, are highlighted as promising avenues to enhance polythene degradation efficiency. The article concludes with a discussion on the potential of microbial perspectives to address plastic pollution and outlines future research directions in this field.

Keywords: Microplastics; degradation; pathways; microbial degradation; environmental impact; plastic pollution; fragmentation; biomass production; mineralization; microorganisms; environmental degradation; biodegradation; enzymatic processes; carbon compounds; sustainable waste management.

1. INTRODUCTION

Plastic pollution has become one of the most pressing environmental challenges of our time, with polythene (polyethylene) contributing significantly to this global crisis. Polythene, renowned for its versatility and durability, is extensively used in various industries, including packaging, construction, and agriculture. However, its non-biodegradable nature poses a severe threat to ecosystems, wildlife, and human health [1-5]. In recent years, the detrimental effects of plastic pollution have spurred intensive research efforts to identify sustainable solutions for plastic waste management. Among these, microbial perspectives on polythene biodegradation have emerged as a promising avenue for mitigating plastic pollution. Microorganisms, including bacteria, fungi, and algae, possess remarkable capabilities to degrade polythene through enzymatic processes and metabolic pathways [6]. AND aims to explore the role of microorganisms in polythene biodegradation and their potential contribution to addressing plastic pollution. We will delve into the mechanisms underlying microbial polythene degradation, including surface erosion, biofilm formation, and metabolic transformations. Furthermore, we will examine the diversity of

polythene-degrading microorganisms found in various environments and the factors influencing their activity and efficiency [7-9].

Understanding microbial perspectives on polythene biodegradation not only sheds light on the natural mechanisms governing plastic degradation but also holds implications for the development of innovative biotechnological solutions. By harnessing the power of microorganisms, we may unlock new strategies for managing plastic waste and preserving the integrity of our environment.

In the following sections, we will explore the intricate relationships between microorganisms and polythene, examine current research findings, and discuss future directions for harnessing microbial perspectives in the fight against plastic pollution [10-15].

2. POLYTHENE BIODEGRADATION MECHANISMS

Polythene biodegradation mechanisms encompass a variety of intricate processes orchestrated by microorganisms. Below, we delve into each mechanism, exploring its underlying principles and implications for plastic pollution mitigation.

2.1 Surface Erosion

Surface erosion represents one of the primary mechanisms by which microorganisms degrade polythene. Enzymatic breakdown of polythene occurs primarily at the surface, where microbial enzymes initiate the degradation process. These extracellular enzymes catalyze chemical reactions that lead to the fragmentation of polythene molecules. Physical and chemical changes induced by microbial activity result in the breakdown of polythene into smaller fragments, eventually rendering it more susceptible to further degradation. Surface erosion plays a crucial role in initiating the degradation of polythene materials in various environmental settings [16,17].

2.2 Biofilm Formation

Microbial biofilms play a pivotal role in enhancing polythene degradation processes. Biofilms are structured microbial communities embedded within a matrix of extracellular polymeric substances (EPS). On polythene surfaces, microorganisms adhere and aggregate to form biofilms, creating a microenvironment conducive to enzymatic activity and metabolic interactions. Within biofilms, microorganisms exhibit increased enzymatic efficiency, allowing for accelerated degradation of polythene materials. The synergistic interactions among biofilm-associated microorganisms further amplify degradation rates, highlighting the importance of microbial community dynamics in plastic biodegradation.

2.3 Metabolic Pathways

Microbial utilization of polythene as a carbon source involves intricate metabolic pathways tailored to degrade complex polymer structures. Polythene-degrading microorganisms possess enzymes capable of breaking down polythene molecules into simpler carbon compounds. These metabolic pathways enable microorganisms to assimilate polythene-derived carbon for energy and growth. Identification and characterization of polythene-degrading enzymes and metabolic intermediates provide insights into the biochemical mechanisms underlying microbial polythene biodegradation.

2.4 Synergistic Interactions

Synergistic interactions among microorganisms drive cooperative behaviors that enhance polythene degradation efficiency. Microbial

communities often consist of diverse species with complementary metabolic capabilities. Through the exchange of metabolites and enzymes, microbial consortia synergistically degrade polythene materials, resulting in more efficient degradation rates than individual microorganisms alone. Synergistic interactions underscore the importance of microbial community composition and diversity in plastic biodegradation processes [18-23].

2.5 Adaptation and Evolution

Microbial adaptation to polythene-rich environments involves genetic and physiological changes that enhance degradation capabilities. Over time, microorganisms evolve specialized enzymes and metabolic pathways optimized for polythene degradation. Through natural selection, microbial populations adapt to environmental pressures imposed by polythene pollution, leading to the emergence of plastic-degrading phenotypes. Understanding microbial adaptation and evolution is critical for predicting the long-term efficacy of microbial-based strategies for plastic pollution mitigation, polythene biodegradation mechanisms involve a complex interplay of enzymatic, metabolic, and ecological processes orchestrated by microorganisms [24-26]. Unraveling the intricacies of these mechanisms holds promise for developing sustainable solutions to combat plastic pollution and preserve environmental health.

1. **Plastic Degradation to Small Fragments:** The process begins with the degradation of plastic waste into smaller fragments through various environmental factors such as UV radiation, mechanical abrasion, and microbial activity. This fragmentation breaks down the plastic into microplastics, which are tiny particles typically less than 5 millimeters in size.
2. **Entry into Cells after Decomposition:** The microplastic fragments, now in smaller sizes, can enter microbial cells through processes such as phagocytosis or passive diffusion. Microorganisms, including bacteria, fungi, and algae, play a crucial role in the degradation of microplastics.
3. **Transformation into Biomass for Energy Production:** Within microbial cells, enzymes and metabolic pathways are activated to degrade the microplastic fragments further. These enzymatic

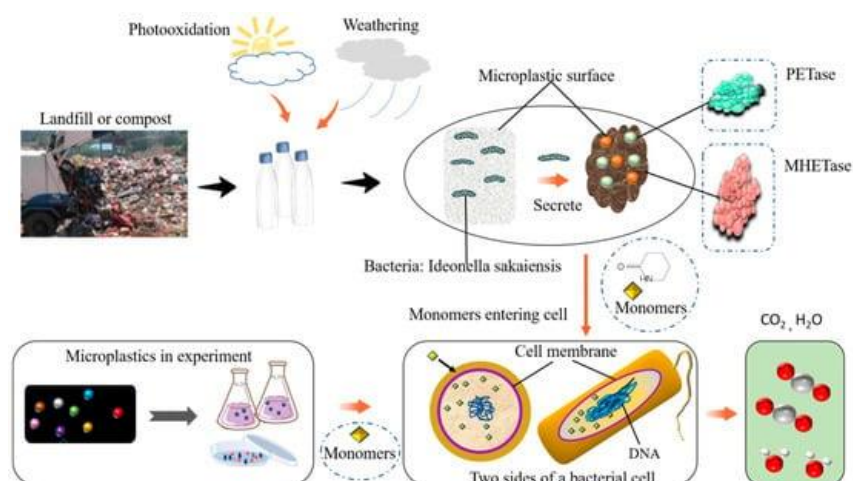


Fig. 1. Illustrates the degradation pathways of microplastics, depicting the journey of plastic fragments as they undergo decomposition and transformation within microbial cells

Source: Cai, Z.; Li, M.; Zhu, Z.; Wang, X.; Huang, Y.; Li, T.; Gong, H.; Yan, M. *Biological Degradation of Plastics and Microplastics: A Recent Perspective on Associated Mechanisms and Influencing Factors*. *Microorganisms* 2023, 11, 1661. <https://doi.org/10.3390/microorganisms11071661>

4. processes break down the chemical bonds of the microplastics, converting them into simpler carbon compounds. These carbon compounds can then be utilized by the microorganisms as a source of energy and building blocks for biomass production [27-30].

Mineralization: Alternatively, the microplastic fragments can undergo mineralization, a process where the carbon compounds derived from the degradation of microplastics are converted into inorganic forms such as carbon dioxide and water. Mineralization effectively returns the carbon back into the environment in a form that can be assimilated by other organisms or sequestered in the ecosystem, the degradation pathways depicted in Fig. 1 highlight the complex interactions between microorganisms and microplastics in the environment. Understanding these pathways is critical for developing strategies to mitigate the impacts of plastic pollution and promote sustainable waste management practices. By elucidating the processes involved in microplastic degradation, researchers can identify potential interventions and technologies to address the growing problem of plastic pollution in our ecosystems.

3. MICROBIAL DIVERSITY AND PLASTIC DEGRADATION

Microbial diversity plays a crucial role in plastic degradation, including the breakdown of polythene (polyethylene). Here, we explore the

relationship between microbial diversity and plastic degradation, highlighting the importance of understanding microbial communities in addressing plastic pollution [31].

3.1 Identification of Polythene-Degrading Microorganisms

Microbial diversity encompasses a wide array of bacteria, fungi, and other microorganisms with the potential to degrade polythene. Research efforts have focused on isolating and characterizing polythene-degrading microorganisms from diverse environments, including soil, water bodies, and waste disposal sites [32-35]. Culture-based and molecular techniques, such as metagenomics and microbial community profiling, have enabled the identification of microbial taxa associated with plastic degradation.

3.2 Exploration of Microbial Communities

Microbial communities associated with plastic degradation exhibit considerable diversity and complexity. Studies have revealed the presence of specialized microbial consortia capable of degrading polythene under various environmental conditions [36-38]. These microbial communities often comprise multiple species with complementary metabolic capabilities, facilitating efficient plastic degradation through synergistic interactions. Understanding the composition and dynamics of microbial communities is essential for elucidating

the mechanisms and environmental factors influencing plastic degradation processes.

3.3 Characterization of Enzymes and Metabolic Pathways

Microbial diversity contributes to the diversity of enzymes and metabolic pathways involved in polythene degradation. Polythene-degrading microorganisms produce a range of enzymes, including lipases, esterases, and oxidases, capable of breaking down polythene molecules into smaller fragments. Metabolic pathways associated with polythene degradation involve the conversion of polythene-derived carbon into metabolic intermediates that can be utilized by microorganisms for energy and growth. Characterizing the enzymatic repertoire and metabolic capabilities of polythene-degrading microorganisms provides insights into the biochemical mechanisms underlying plastic degradation [39-48].

3.4 Environmental Factors Influencing Microbial Diversity and Plastic Degradation

Environmental factors, such as temperature, pH, moisture, and nutrient availability, play significant roles in shaping microbial diversity and plastic degradation processes. Microbial communities exhibit varying degrees of plastic degradation activity in response to environmental conditions, highlighting the importance of optimizing environmental parameters for enhanced plastic degradation efficiency. Additionally, anthropogenic factors, including pollution levels and habitat disturbance, can influence microbial community composition and plastic degradation rates [49].

3.5 Implications for Plastic Waste Management

Understanding microbial diversity and plastic degradation mechanisms has profound implications for plastic waste management strategies. Harnessing the metabolic capabilities of diverse microbial communities offers potential solutions for mitigating plastic pollution and promoting environmental sustainability. Biotechnological approaches, such as the development of microbial consortia and genetic engineering of polythene-degrading microorganisms, hold promise for enhancing plastic degradation efficiency and scalability [50],

microbial diversity plays a pivotal role in plastic degradation, offering insights into the complex interactions between microorganisms and polythene. By elucidating the mechanisms and environmental factors influencing plastic degradation processes, researchers can develop innovative strategies for addressing plastic pollution and advancing sustainable waste management practices.

Biofilm formation represents a key mechanism through which microbial communities enhance degradation processes, including the breakdown of polythene and other environmental pollutants. In biofilms, microorganisms aggregate and adhere to surfaces, encapsulating themselves within a matrix of extracellular polymeric substances (EPS). This matrix provides structural support and protection to the microbial community, facilitating their survival and metabolic activities in challenging environmental conditions [51-53].

Biofilms play a pivotal role in enhancing degradation processes through several mechanisms:

1. **Surface Adhesion and Colonization:** Microorganisms within biofilms adhere to polythene surfaces, enabling them to colonize and establish stable communities. Adhesion is mediated by microbial surface structures and adhesive molecules, allowing for close proximity to polythene substrates.
2. **Microbial Cooperation and Synergy:** Within biofilms, microbial communities engage in cooperative behaviors and metabolic synergy. Different microbial species within the biofilm interact through metabolic exchange, sharing nutrients, signaling molecules, and metabolic by-products. This cooperative interaction enhances the overall degradation efficiency of the biofilm community.
3. **Localized Microenvironments:** Biofilms create localized microenvironments that support specialized metabolic activities. Variations in nutrient availability, pH, oxygen levels, and other environmental factors occur within biofilms, leading to the emergence of distinct metabolic niches. These microenvironments can promote specific enzymatic activities and metabolic pathways involved in polythene degradation.
4. **Protection from Environmental Stressors:** The EPS matrix surrounding biofilm cells

5. provides protection against environmental stressors, including UV radiation, desiccation, and chemical toxins. The matrix acts as a barrier, shielding microbial cells from harmful agents while retaining essential nutrients and moisture necessary for metabolic activity.

Retention of Enzymes and Metabolites: Enzymes and metabolites produced by microbial cells within biofilms are retained and concentrated within the EPS matrix. This spatial organization facilitates the accumulation of enzymatic activity at polythene surfaces, enhancing degradation efficiency and substrate accessibility.

Overall, biofilm formation represents a sophisticated strategy employed by microbial communities to enhance degradation processes in diverse environmental contexts. Understanding the dynamics of biofilm-mediated degradation is essential for harnessing the potential of microbial communities in bioremediation and sustainable waste management strategies, including the mitigation of plastic pollution [37,38,51].

4. ENVIRONMENTAL FACTORS INFLUENCING POLYTHENE BIODEGRADATION

Environmental factors significantly influence polythene biodegradation, shaping the activity and efficiency of microbial degradation processes. Understanding these factors is crucial for optimizing conditions to enhance polythene degradation rates and explore key environmental factors that influence polythene biodegradation:

4.1 Temperature

- Temperature exerts a profound influence on microbial activity and enzymatic processes involved in polythene degradation.
- Generally, higher temperatures accelerate microbial metabolism and enzymatic activity, leading to increased degradation rates.
- However, extreme temperatures can denature enzymes and disrupt microbial activity, affecting degradation efficiency.

4.2 pH

- pH levels influence enzyme activity and microbial growth, thus impacting polythene degradation.

- Optimal pH ranges vary depending on the microbial species and enzymes involved in polythene degradation.
- Fluctuations in pH levels can alter microbial community composition and enzymatic activity, affecting degradation kinetics.

4.3 Moisture Content

- Adequate moisture is essential for microbial growth, enzyme function, and polythene hydrolysis.
- Moisture levels influence microbial colonization and biofilm formation on polythene surfaces, facilitating degradation processes.
- Excessive moisture may lead to waterlogging and oxygen depletion, impeding microbial activity and degradation efficiency.

4.4 Nutrient Availability

- Microbial degradation of polythene requires essential nutrients, including carbon, nitrogen, phosphorus, and trace elements.
- Imbalances in nutrient availability can limit microbial growth and metabolic activity, thereby affecting degradation rates.
- Nutrient supplementation or organic amendments may enhance microbial activity and polythene degradation in nutrient-poor environments.

4.5 Oxygen Availability

- Aerobic conditions favor microbial polythene degradation, as oxygen serves as a terminal electron acceptor in aerobic respiration.
- Aerobic microorganisms produce more energy-efficient metabolic pathways for polythene degradation compared to anaerobic microorganisms.
- Oxygen depletion in anaerobic environments may slow down polythene degradation rates and favor the accumulation of recalcitrant degradation by-products.

4.6 Substrate Characteristics

- Physical and chemical properties of polythene substrates, such as surface

area, crystallinity, molecular weight, and additives, influence degradation kinetics.

- Microorganisms exhibit substrate specificity, with certain strains preferentially degrading specific types of polythene polymers.
- Surface modifications, such as roughening or chemical treatments, can enhance microbial attachment and polythene degradation efficiency.

Optimizing environmental conditions based on these factors can enhance microbial polythene degradation rates and promote more sustainable waste management practices. Additionally, understanding the interplay between environmental factors and microbial activity is essential for developing effective bioremediation strategies to mitigate plastic pollution [54,55].

5. BIOTECHNOLOGICAL APPROACHES AND FUTURE DIRECTIONS

Biotechnological approaches offer innovative strategies for enhancing polythene biodegradation and addressing plastic pollution. These approaches leverage advances in microbiology, biotechnology, and genetic engineering to develop novel solutions for plastic waste management. Here, we discuss biotechnological approaches and future directions in the field of polythene biodegradation:

5.1 Microbial Consortia Development

- Engineering microbial consortia comprising diverse polythene-degrading microorganisms can synergistically enhance degradation efficiency.
- By combining microorganisms with complementary metabolic capabilities, microbial consortia can target different stages of polythene degradation and improve overall degradation rates.
- Optimization of microbial consortia composition and environmental conditions can maximize polythene degradation efficiency and stability.

5.2 Genetic Engineering of Microorganisms

- Genetic engineering enables the modification of microbial strains to

enhance their polythene-degrading capabilities.

- Engineering microorganisms to overexpress key enzymes involved in polythene degradation pathways can accelerate degradation rates and improve substrate specificity.
- Synthetic biology approaches facilitate the design and construction of microbial biosystems optimized for polythene degradation under diverse environmental conditions.

5.3 Enzyme Engineering and Protein Engineering

- Enzyme engineering techniques, such as directed evolution and rational design, can enhance the catalytic efficiency and substrate specificity of polythene-degrading enzymes.
- Protein engineering strategies enable the modification of enzyme structures to improve stability, activity, and compatibility with polythene substrates.
- Engineered enzymes can be deployed in bioreactor systems or enzyme cocktails for efficient polythene degradation in industrial and environmental settings.

5.4 Bioinformatics and Metagenomics

- Bioinformatics tools and metagenomic approaches facilitate the discovery and characterization of novel polythene-degrading enzymes and metabolic pathways.
- Metagenomic analysis of environmental microbial communities can uncover genetic resources for polythene degradation and identify candidate enzymes with unique catalytic properties.
- Integration of bioinformatics data with experimental validation enables the identification of microbial consortia and enzymes with potential applications in polythene biodegradation.

5.5 Synthetic Biology and Systems Biology

- Synthetic biology platforms enable the design and construction of synthetic genetic circuits and microbial biosystems for targeted polythene degradation. Systems biology approaches provide

insights into the dynamic interactions between microbial communities and polythene substrates, elucidating complex degradation pathways and regulatory networks. Integration of synthetic biology and systems biology methodologies offers a holistic understanding of microbial polythene degradation and enables the engineering of tailored solutions for plastic waste management, biotechnological approaches hold immense potential for revolutionizing polythene biodegradation and addressing plastic pollution on a global scale. Continued research efforts and interdisciplinary collaborations will drive innovation in the field, paving the way for sustainable solutions to the challenges posed by plastic waste in the environment [56].

6. CONCLUSION

In conclusion, the exploration of microbial perspectives on polythene biodegradation represents a promising avenue for addressing the pervasive problem of plastic pollution. Throughout this discourse, we have delved into the intricate mechanisms by which microorganisms degrade polythene, highlighting the diverse enzymatic, metabolic, and ecological processes involved in plastic degradation.

Microorganisms possess remarkable capabilities to degrade polythene through surface erosion, biofilm formation, metabolic pathways, synergistic interactions, and adaptation to environmental conditions. The identification of polythene-degrading microorganisms and the characterization of their enzymatic repertoire provide insights into the biochemical mechanisms underlying plastic degradation.

Environmental factors such as temperature, pH, moisture content, nutrient availability, and oxygen levels significantly influence polythene biodegradation processes. Optimization of environmental conditions and the development of biotechnological approaches, including microbial consortia development, genetic engineering, enzyme engineering, and synthetic biology, hold promise for enhancing polythene degradation efficiency and scalability. Furthermore, the exploration of microbial diversity and plastic degradation mechanisms offers valuable insights into the complex interactions between microorganisms and polythene substrates. Harnessing the metabolic capabilities of diverse

microbial communities provides innovative strategies for mitigating plastic pollution and promoting environmental sustainability, interdisciplinary collaborations, technological innovations, and policy initiatives will be essential for advancing research in microbial polythene biodegradation and translating scientific knowledge into practical solutions for plastic waste management. By leveraging microbial perspectives and biotechnological approaches, we can work towards a future where plastic pollution is effectively mitigated, and the integrity of our environment is preserved for generations to come.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Danso D, Chow J, Streit WR. Plastics: Environmental and biotechnological perspectives on microbial degradation. *Applied and Environmental Microbiology*. 2019;85(19):e01095-19.
2. Liaqat S, Hussain M, Malik MF, Aslam A, Mumtaz K. Microbial ecology: A new perspective of plastic degradation. *Pure and Applied Biology*. 2020;9(4):2138-2150.
3. Purohit J, Chattopadhyay A, Teli B. Metagenomic exploration of plastic degrading microbes for biotechnological application. *Current Genomics*. 2020;21(4):253-270.
4. Cf SF, Rebello S, Mathachan Aneesh E, Sindhu R, Binod P, Singh S, Pandey A. Bioprospecting of gut microflora for plastic biodegradation. *Bioengineered*. 2021;12(1):1040-1053.
5. Oliveira J, Belchior A, da Silva VD, Rotter A, Petrovski Ž, Almeida PL, Gaudêncio SP. Marine environmental plastic pollution: Mitigation by microorganism degradation and recycling valorization. *Frontiers in Marine Science*. 2020;7:567126.
6. Carr CM, Clarke DJ, Dobson AD. Microbial polyethylene terephthalate hydrolases: Current and future perspectives. *Frontiers in Microbiology*. 2020;11:571265.
7. Brueckner T, Eberl A, Heumann S, Rabe M, Guebitz GM. Enzymatic and chemical hydrolysis of poly (ethylene terephthalate) fabrics. *J. Polym. Sci. Part A* 46. 2008;6435–6443. DOI: 10.1002/pola.22952

8. Cam Y, Alkim C, Trichez D, Trebosc V, Vax A, Bartolo F, et al. Engineering of a synthetic metabolic pathway for the assimilation of (d)-Xylose into value-added chemicals. *ACS Synth. Biol.* 2016;5:607–618.
DOI: 10.1021/acssynbio.5b00103
9. Carniel A, Valoni E, Nicomedes J, Gomes ADC, Castro AMD. Lipase from *Candida antarctica* (CALB) and cutinase from *Humicola insolens* act synergistically for PET hydrolysis to terephthalic acid. *Process Biochem.* 2017;59:84–90.
DOI: 10.1016/j.procbio.2016.07.023
10. Carta D, Cao G, D'Angeli C. Chemical recycling of poly(ethylene terephthalate) (pet) by hydrolysis and glycolysis. *Environ. Sci. Pollut. Res.* 2003;10:390–394.
DOI: 10.1065/espr2001.12.104.8
11. Chen CC, Han X, Ko TP, Liu W, Guo RT. Structural studies reveal the molecular mechanism of PETase. *FEBS J.* 2018; 285:3717–3723.
DOI: 10.1111/febs.14612
12. Chen S, Su L, Chen J, Wu J. Cutinase: Characteristics, preparation, and application. *Biotechnol. Adv.* 2013;31: 1754–1767.
DOI: 10.1016/j.biotechadv.2013.09.005
13. Chen S, Tong X, Woodard RW, Du G, Wu J, Chen J. Identification and characterization of bacterial cutinase. *J. Biol. Chem.* 2008;283:25854–25862.
DOI: 10.1074/jbc.M800848200
14. Chen Z, Wang Y, Cheng Y, Wang X, Tong S, Yang H, et al. Efficient biodegradation of highly crystallized polyethylene terephthalate through cell surface display of bacterial PETase. *Sci. Total Environ.* 2020;709:136138.
DOI: 10.1016/j.scitotenv.2019.136138
15. Child J, Willetts A. Microbial metabolism of aliphatic glycols bacterial metabolism of ethylene glycol. *Biochim. Biophys. Acta.* 1978;538:316–327.
DOI: 10.1016/0304-4165(78)90359-8
16. da Costa JP, Mouneyrac C, Costa M, Duarte AC, Rocha-Santos T. The role of legislation, regulatory initiatives and guidelines on the control of plastic pollution. *Front. Environ. Sci.* 2020;8:104.
DOI: 10.3389/fenvs.2020.00104
17. Danso D, Chow J, Streit WR. Plastics: Microbial degradation, environmental and biotechnological perspectives. *Appl. Environ Microbiol.* 2019;85:AEM.01095–1019.
18. Gong J, Duan N, Zhao X. Evolutionary engineering of *Phaffia rhodozyma* for astaxanthin-overproducing strain. *Front. Chem. Sci. Eng.* 2012;6:174–178.
DOI: 10.1007/s11705-012-1276-3
19. Gong J, Kong T, Li Y, Li Q, Li Z, Zhang J. Biodegradation of microplastic derived from poly (ethylene terephthalate) with bacterial whole-cell biocatalysts. *Polymers.* 2018;10:1326.
DOI: 10.3390/polym10121326
20. Sangameshwar R, Rasool A, Venkateshwar, C. (2020). Effect of heavy metals on leafy vegetable (*Trigonella foenum-graecum* L.) and its remediation. *Plant Archives*, 20(2), 1941-1944.
21. Griswold KE, Mahmood NA, Iverson BL, Georgiou G. Effects of codon usage versus putative 5'-mRNA structure on the expression of *Fusarium solani* cutinase in the *Escherichia coli* cytoplasm. *Protein Expr. Purif.* 2003;27:134–142.
DOI: 10.1016/S1046-5928(02)00578-8
22. Groeninckx G, Berghmans H, Overbergh N, Smets G. Crystallization of poly (ethylene terephthalate) induced by inorganic compounds. I. Crystallization behavior from the glassy state in a low-temperature region. *J. Polym. Sci.* 1974;12:303–316.
DOI: 10.1002/pol.1974.180120207
23. Guebitz GM, Cavaco-Paulo A. Enzymes go big: Surface hydrolysis and functionalisation of synthetic polymers. *Trends Biotechnol.* 2008;26:32–38.
DOI: 10.1016/j.tibtech.2007.10.003
24. Danso D, Schmeisser C, Chow J, Zimmermann W, Wei R, Leggewie C, et al. New insights into the function and global distribution of polyethylene terephthalate (PET)-degrading bacteria and enzymes in marine and terrestrial metagenomes. *Appl. Environ. Microbiol.* 2018;84:e02773-17.
25. Demirel B, Yaraş A, Elçiçek H. Crystallization behavior of PET materials. *BAÜ Fen. Bil. Enst. Dergisi. Cilt.* 2011;13:26–35.
26. Dodd D, Mackie RI, Cann IK. Xylan degradation, a metabolic property shared by rumen and human colonic Bacteroidetes. *Mol. Microbiol.* 2011;79:292–304.
DOI: 10.1111/j.1365-2958.2010.07473.x
27. Hahladakis JN, Iacovidou E, Gerassimidou S. Chapter 19 - plastic waste in a circular

- economy. in plastic waste and recycling, ed. T. M. Letcher (Cambridge, MA: Academic Press). 2020;481–512. DOI: 10.1016/b978-0-12-817880-5.00019-0
28. Rasool A, Mir MI, Zulfajri M, Hanafiah MM, Unnisa SA, Mahboob M. Plant growth promoting and antifungal asset of indigenous rhizobacteria secluded from saffron (*Crocus sativus* L.) rhizosphere. *Microbial Pathogenesis*. 2021;150:104734.
 29. Hahladakis JN, Velis CA, Weber R, Iacovidou E, Purnell P. An overview of chemical additives present in plastics: Migration, release, fate and environmental impact during their use, disposal and recycling. *J. Hazard. Mater*. 2018;344:179–199. DOI: 10.1016/j.jhazmat.2017.10.014
 30. Sultana N, Saini PK, Kiran SR, Kanaka S. Exploring the antioxidant potential of medicinal plant species: A comprehensive review. *Journal of Plant Biota*; 2023.
 31. Donelli I, Freddi G, Nierstrasz VA, Taddei P. Surface structure and properties of poly-(ethylene terephthalate) hydrolyzed by alkali and cutinase. *Polym. Degrad. Stab*. 2010;95:1542–1550. DOI:10.1016/j.polymdegradstab.2010.06.011
 32. Eberl A, Heumann S, Brückner T, Araujo R, Cavaco-Paulo A, Kaufmann F, et al. Enzymatic surface hydrolysis of poly (ethylene terephthalate) and bis(benzoyloxyethyl) terephthalate by lipase and cutinase in the presence of surface active molecules. *J. Biotechnol*. 2009;143:207–212. DOI: 10.1016/j.jbiotec.2009.07.008
 33. Elangovan S, Pandian SBS, Geetha S, Joshi SJ. Polychlorinated Biphenyls (PCBs): Environmental fate, challenges and bioremediation. in *Microbial Metabolism of Xenobiotic Compounds*. New York, NY: Springer. 2019;165–188.
 34. Farzi A, Dehnad A, Fotouhi AF. Biodegradation of polyethylene terephthalate waste using *Streptomyces* species and kinetic modeling of the process. *Biocatal. Agricult. Biotechnol*. 2019;17:25–31. DOI: 10.1016/j.bcab.2018.11.002
 35. Fecker T, Galaz-Davison P, Engelberger F, Narui Y, Sotomayor M, Parra LP, et al. Active site flexibility as a hallmark for efficient PET degradation by *I. sakaiensis* PETase. *Biophys. J*. 2018;114:1302–1312. DOI: 10.1016/j.bpj.2018.02.005
 36. Franden MA, Jayakody LN, Li W-J, Wagner NJ, Cleveland NS, Michener WE, et al. Engineering *Pseudomonas putida* KT2440 for efficient ethylene glycol utilization. *Metab. Eng*. 2018;48:197–207. DOI: 10.1016/j.ymben.2018.06.003
 37. Frazee RW, Livingston DM, LaPorte DC, Lipscomb JD. Cloning, sequencing, and expression of the *Pseudomonas putida* protocatechuate 3, 4-dioxygenase genes. *J. Bacteriol*. 1993;175: 6194–6202. DOI: 10.1128/jb.175.19.6194-6202.1993
 38. Furukawa M, Kawakami N, Oda K, Miyamoto K. Acceleration of enzymatic degradation of poly (ethylene terephthalate) by surface coating with anionic surfactants. *Chem Sus Chem*. 2018; 11: 4018–4025. DOI: 10.1002/cssc.201802096
 39. Hajighasemi M, Tchigvintsev A, Nocek B, Flick R, Popovic A, Hai T, et al. Screening and characterization of novel polyesterases from environmental metagenomes with high hydrolytic activity against synthetic polyesters. *Environ. Sci. Technol*. 2018;52:12388–12401. DOI: 10.1021/acs.est.8b04252
 40. Han X, Liu W, Huang J-W, Ma J, Zheng Y, Ko TP, et al. Structural insight into catalytic mechanism of PET hydrolase. *Nat. Commun*. 2017;8:2106.
 41. Handelsman J. Metagenomics: Application of genomics to uncultured microorganisms. *Microbiol. Mol. Biol. Rev*. 2004;68:669–685. DOI: 10.1128/MMBR.68.4.669-685.2004
 42. Herrero Acero E, Ribitsch D, Dellacher A, Zitzenbacher S, Marold A, Steinkellner G, et al. Surface engineering of a cutinase from *Thermobifida cellulolytica* for improved polyester hydrolysis. *Biotechnol. Bioeng*. 2013;110:2581–2590. DOI: 10.1002/bit.24930
 43. Herrero Acero E, Ribitsch D, Steinkellner G, Gruber K, Greimel K, Eiteljoerg I, et al. Enzymatic surface hydrolysis of PET: Effect of structural diversity on kinetic properties of cutinases from *Thermobifida*. *Macromolecules*. 2011;44:4632–4640. DOI: 10.1021/ma200949p
 44. Rasool A, Kanagaraj T, Mir MI, Zulfajri M, Ponnusamy VK, Mahboob M. Green coalescence of CuO nanospheres for efficient anti-microbial and anti-cancer conceivable activity. *Biochemical Engineering Journal*. 2022;187:108464.

45. Hiraga K, Taniguchi I, Yoshida S, Kimura Y, Oda K. Biodegradation of waste PET. EMO Rep. 2019;20:e49365.
46. Hosaka M, Kamimura N, Toribami S, Mori K, Kasai D, Fukuda M, et al. Novel tripartite aromatic acid transporter essential for terephthalate uptake in *Comamonas* sp. strain E6. Appl. Environ. Microbiol. 2013;79:6148–6155. DOI: 10.1128/aem.01600-13
47. Arpana, K K Gill, Samanpreet Kaur, Kavita Bhatt and S S Sandhu. Decadal Analysis of Rainy Days and Extreme Rainfall Events in Different Agroclimatic Zones of Punjab. Agriculture Archives; 2024. DOI:https://doi.org/10.51470/AGRI.2024.3.1.01
48. Huang X, Cao L, Qin Z, Li S, Kong W, Liu Y. Tat-independent secretion of polyethylene terephthalate hydrolase PETase in *Bacillus subtilis* 168 mediated by its native signal peptide. J. Agricult. Food Chem. 2018;66:13217–13227. DOI: 10.1021/acs.jafc.8b05038
49. Igiri BE, Okoduwa SI, Idoko GO, Akabuogu EP, Adeyi AO, Ejiogu IK. Toxicity and bioremediation of heavy metals contaminated ecosystem from tannery wastewater: A review. J. Toxicol. 2018; 2568038.
50. Jaeger K-E, Ransac S, Dijkstra BW, Colson C, van Heuvel M, Misset O. Bacterial lipases. FEMS Microbiol. Rev. 1994;15:29–63.
51. Furukawa M, Kawakami N, Tomizawa A, Miyamoto K. Efficient degradation of poly(ethylene terephthalate) with thermobifida fusca cutinase exhibiting improved catalytic activity generated using mutagenesis and additive-based approaches. Sci. Rep. 2019;9:16038. DOI: 10.1038/s41598-019-52379-z
52. Gamerith C, Zartl B, Pellis A, Guillamot F, Marty A, Acero EH, et al. Enzymatic recovery of polyester building blocks from polymer blends. Process Biochem. 2017; 59:58–64. DOI: 10.1016/j.procbio.2017.01.004
53. Garside M. Global PET bottle production 2004-2021; 2019. Available: <https://www.statista.com/statistics/723191/production-of-polyethylene-terephthalate-bottles-worldwide/> Accessed on: October 04, 2020.
54. Jaya Prabhavathi S, Subrahmanian K, Senthil Kumar M, Gayathry G, Malathi G. Exploring the Antibacterial, Anti-Biofilm, and Anti-Quorum Sensing Properties of Honey: A Comprehensive Review. Agriculture Archives; 2023. DOI: <https://doi.org/10.51470/AGRI.2023.2.3.10>
55. Islam MS, Rahman MM, Paul NK. Arsenic-induced morphological variations and the role of phosphorus in alleviating arsenic toxicity in rice (*Oryza sativa* L.). Plant Science Archives. 2016;1(1):1-10.
56. Niranjana C. Characterization of bacteriocin from lactic acid bacteria and its antibacterial activity against *Ralstonia solanacearum* causing tomato wilt. Plant Science Archives; 2016.

© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/113576>