

Journal of Advances in Medical and Pharmaceutical Sciences

Volume 25, Issue 8, Page 1-11, 2023; Article no.JAMPS.104196 ISSN: 2394-1111

# Pharmacological Investigation of *Invitro* Anti-inflammatory, Antimicrobial, Thrombolytic, Cytotoxic and *In vivo* Analgesic Activities of Ethanolic Leaf Extract of *Diospyros malabarica*

Maria Chowdhury <sup>a++</sup>, Lamia Akter Sultana <sup>a#</sup>, Azijun Chowdhury Joya <sup>a#</sup> and Hasib Khan Shomudro <sup>b†\*</sup>

> <sup>a</sup> Department of Pharmacy, University of Asia Pacific, Dhaka, Bangladesh. <sup>b</sup> Department of Pharmacy, Stamford University Bangladesh, Dhaka, Bangladesh.

## Authors' contributions

This work was carried out in collaboration among all authors. All authors contributed to the completion of this research work. Author MC carried out the laboratory work per the procedure with her co authors LAS and ACJ. Author MC conducted a literature search and statistical analysis. Authors HKS and MC designed the research and wrote the draft manuscript. Author HKS supervised the whole research project. All authors read and approved the final manuscript.

## Article Information

DOI: 10.9734/JAMPS/2023/v25i8630

**Open Peer Review History:** 

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/104196

> Received: 20/06/2023 Accepted: 23/08/2023 Published: 28/08/2023

**Original Research Article** 

++Student of Pharmacy (Masters);

<sup>&</sup>lt;sup>#</sup> Student of Pharmacy (Honors);

<sup>&</sup>lt;sup>†</sup> Student, Bachelor of Pharmacy (Honors);

<sup>\*</sup>Corresponding author: E-mail: hasibkhan.shomudro@hotmail.com;

J. Adv. Med. Pharm. Sci., vol. 25, no. 8, pp. 1-11, 2023

## ABSTRACT

**Aims:** This study was designed to investigate the effects of leaf extracts of *Diospyros malabarica* plant (EEDML) on a number of in vitro activities, including antibacterial, anti-inflammatory, cytotoxic, and thrombolytic activities, as well as in vivo analgesic activities.

**Place and Duration of Study:** The Laboratory of Phytochemistry and Pharmacology at the Department of Pharmacy and the Laboratory of Microbiology at Stamford University Bangladesh and the Bangladesh Council of Scientific and Industrial Research, Dhaka, conducted this investigation from January 2022 to June 2023.

**Methodology:** The study employed the Egg Albumin Protein Denaturation Assay to evaluate the anti-inflammatory effects of various doses. The Disk Diffusion Method was employed for conducting an antimicrobial assay. Different tests were conducted to evaluate the effects of thrombolytic and cytotoxic substances. These tests included the Clot Lysis assay and the Brine Shrimp Lethality Assay. In-vivo analgesic testing was conducted using the hot plate method and the acetic acid-induced writhing test.

**Results:** The experimental tests showed significant pain-relieving effects, with inhibition percentages of 46.2% and 66.61% observed at doses of 200 and 400 mg/kg, respectively. The anti-inflammatory test showed a significant inhibition of EEDML at a concentration of 1000 $\mu$ g/mL, with an inhibition rate of 85.45%. The antimicrobial test showed a moderate antimicrobial effect, with a zone of inhibition ranging from 07-15, depending on the microorganism and dosage. The thrombolytic test showed a clot lysis rate of 85.37%, while the cytotoxic test indicated an LC50 value of 1.47.

**Conclusion:** EEDML's phytochemical displays strong pharmacological properties, suggesting its potential for drug discovery in diverse areas.

Keywords: Anti-inflammatory; antimicrobial; cytotoxic; thrombolytic; analgesic.

## 1. INTRODUCTION

The utilization of medicinal plants has been a significant contributor to the advancement of healthcare in contemporary times. Natural compounds with therapeutic properties have been identified as a valuable source for the development of new drugs. These compounds have been found to serve as a foundation for the creation of novel therapeutic agents. In a study published in Phytomedicine, it was reported that around 25% of prescription drugs worldwide are derived from plants or plant-based compounds [1]. For example, Paclitaxel, a chemotherapeutic agent, has been derived from the Pacific yew tree (Taxus brevifolia) and has demonstrated efficacy against a range of cancers [2]. Artemisinin, a sesquiterpene lactone, is an essential constituent in the treatment and management of malaria. It is extracted from the plant Artemisia annua, commonly known as sweet wormwood. The significance of artemisinin in the treatment of malaria has been well documented [3]. The utilization of medicinal plants has been recognized as a promising avenue for drug discovery and development due to their abundant bioactive compounds. The opium poppy (Papaver somniferum) is known to contain alkaloids that exhibit strong analgesic properties [4]. One such alkaloid is morphine. The presence of terpenoids in various plant species, such as Panax ginseng and Curcuma longa, has been observed to demonstrate significant anti-inflammatory and antioxidant properties [5]. In addition, the potential of flavonoids, which are found in abundance in fruits and vegetables, has been investigated in relation to their anticancer, anti-inflammatory, and neuroprotective effects [6,7]. The use of Ayurvedic herbs has gained significant popularity due to their various therapeutic effects. Among these herbs, Ashwagandha (Withania somnifera) and Turmeric have been extensively studied for potential health benefits their [8]. The significance of medicinal plants in the contemporary era is noteworthy due to their potential to stimulate novel drug development. valuable furnish natural compounds. and amalgamate traditional medicine with conventional healthcare practices. In light of these factors, it can be concluded that medicinal plants hold a pivotal position in the field of medicine.

*Diospyros malabarica* is a botanical species classified as a flowering tree, belonging to the

family Ebenaceae. It is predominantly found in the South-East region of Asia, where it is native to the local flora. The utilization of fruit extract as a therapeutic agent for the treatment of various ailments, including diabetes, dates back to ancient times [9]. This evergreen tree has the potential to reach a height of 25 meters and has a crown that is densely packed with glossy, dark green leaves. Diospyros malabarica, commonly known as the Malabar persimmon, produces diminutive, spheroidal drupes that undergo a color transformation from green to yellow or orange upon reaching maturity. They possess the quality of edibility and are characterized by a taste profile that is both sweet and tangy. The consumption of fruits is widely accepted in the surrounding region, and they are also utilized in traditional medicine due to their diverse properties. Ethanolic extract medicinal of Diospyros malabarica leaf (EEDML) is associated with antioxidant, antimicrobial, and anti-inflammatory properties [10]. According to [11] the plant Diospyros malabarica is purported to possess properties that exhibit anti-diabetic, anti-inflammatory, and anti-ulcer effects. The present study focuses on the medicinal properties of Diospyros malabarica leaves. The leaves of this plant have been traditionally used for treating various ailments such as skin diseases, rheumatism, and digestive disorders.

In the current research, the organic soluble components of a methanol extract of the complete plant were examined for the first time for their in-vitro anti-inflammatory, antimicrobial, thrombolytic, cytotoxic and in vivo analgesic activity. These activities were investigated for the ethanolic leaf extract of the plant

## 2. MATERIALS AND METHODS

## 2.1 Sample Collection

In Monohorganj, Cumilla, Bangladesh, a leaf of the *Diospyros malabarica* plant was harvested in July 2022. The specimens were brought in for identification by a Scientific Officer from the University of Dhaka's Botany Department in Bangladesh. For the purpose of research, it has been preserved by submitting a voucher specimen (10815) in the Herbarium.

## 2.2 Plant Extract Preparation

The plant leaves were prepared for usage after three weeks of air drying at room temperature. The plant parts were dried, ground into powder, sieved, and then kept in plastic containers. A total of 874g of the dried plant was pulverized and then percolated in 1000 mL of ethanol for 24 hours. The extract was then filtered using filtration method with filter papers before being placed in a conical flask. The resultant extracts were concentrated and dried out with a rotary evaporator [12].

## 2.3 Reagents

Sigma Chemical Co., USA provided methanol, NaOH, diluted HCl acid, concentrated H<sub>2</sub>SO<sub>4</sub>. The sterile saline solution was purchased from Orion Infusion Ltd. Diclofenac sodium was produced by Square Pharmaceuticals Ltd. Vincristine sulphate was purchased from Polysciences, Inc. India. Streptokinase was purchased from Incepta Pharmaceuticals Ltd, Bangladesh. Vin-Cristine Sulphate was sourced from Celon Laboratories Pvt. Ltd. while, Gonoshasthaya Pharmaceuticals Ltd. provided morphine sulfate.

## 2.4 *In-vitro* Anti-inflammatory Test and Protocol

The experimental solution was prepared by combining 0.2 mL of egg albumin obtained from a fresh hen's egg, 2.8 mL of phosphatebuffered saline (PBS) with a pH of 6.4, and 2 mL of an extract with varying concentrations. The final concentrations of the extract were adjusted to 100, 200, 300, 400, and 500 µg/mL. The total volume of the reaction mixture was 5 mL. A control was utilized in the experiment, consisting of an equivalent volume of double-distilled water. Subsequently, the mixtures were subjected to incubation at a temperature of (37°C ± 2) within a BOD incubator manufactured by Lab-Line Technologies for a duration of 15 minutes, followed by a heating process at 70°C for a period of 5 minutes. Following the cooling process, the absorbance of the sample was assessed at a wavelength of 660 nm (utilizing a SHIMADZU UV 1800 instrument). The reference drug, Acetyl Salicylic Acid, was utilized at varying final concentrations of 100,200, 300, 400, and 500µg/mL. The drug was subjected to a similar treatment for the purpose of absorbance determination as described by [13]. The formula for determining the proportion of protein denaturation that was prevented is as follows:

 $\frac{(Absorbance of control-Absorbance of sample)}{Absorbance of control} \times 100$ 

#### **2.5 Antimicrobial Test**

#### 2.5.1 Sources of microorganisms

Pure culture of fungi (Penicillium chrvsogenum. Aspergillus niger, Mucor hiemalis and Yeast budding) and pure culture of Gram-positive (Staphylococcus aureus, Bacillus megaterium) and Gram-negative (Klebsiella pneumonia, Escherichia coli, Pseudomonas aeruginosa) bacterial pathogens were obtained from Microbiology Department of Stamford University Bangladesh and Bangladesh Council of Scientific and Industrial Research.

#### 2.5.2 Antimicrobial susceptibility test

In this research, sterile discs with a diameter of 6.0 mm obtained from Becton Dickinson Microbiology System in the United States were utilized to impregnate three distinct dilutions of the extracts. The extract is available in concentrations of 30, 50, and 70 mg/mL.

The discs were subjected to a storage temperature of -5°C prior to their utilization. The disc diffusion method was employed to conduct the tests. Impregnated discs were positioned on agar and subjected to incubation at either 37°C for a duration of 24-48 hours for bacterial growth or at 30°C for a period of 72 hours for fungal growth. The presence of clear zones of inhibition was utilized to determine the antibacterial or antifungal activities [14].

#### 2.6 Thrombolytic Test

#### 2.6.1 Blood specimen

Ten (10) human volunteers who had no history of oral contraceptive or blood thinner use were selected, and 4 mL of whole blood was collected from a vein.

According to the protocol sanctioned by the Institutional Ethics Council of Stamford University Bangladesh. A phlebotomist assisted venous blood sample collection. Following the collection of the blood sample, 500  $\mu$ L of the sample was dispensed into each of the 10 microcentrifuge tubes.

## 2.6.2 Statement on informed consent of the donors

The investigators provided the volunteer participants with a consent form that explicated the purpose of the inquiry, along with the

appellation of the research project, the identities of the investigators, and their contact particulars. The research has been carried OUIT comprehensive account detailing the criteria for inclusion and exclusion of donors. the administration or non-administration of therapy to donors, the quantity of blood to be extracted, the probable discomfort of the puncture sites, and the duration of time required for blood sampling.

#### 2.6.3 Thrombolysis

The method used for clot lysis experiments was conducted in accordance with the procedures outlined in a previously published research article [15], whereby the percentage of total clot lysis was expressed. To summarize, a total of 10 sterile microcentrifuge tubes, each pre-weighed and with a volume of 0.5 mL, were utilized to contain 2.5 mL of venous blood obtained from healthy subjects. The tubes were then subjected to incubation at a temperature of 37°C for a duration of 45 minutes. Following the formation of a clot, the serum was extracted from the tubes while ensurina that the clot remained undisturbed. The tubes were subsequently reweighed to determine the weight of the clot, which was calculated by subtracting the weight of the tube from the weight of the tube containing the clot. To ensure proper supplementation, it is necessary to add 100 µL of EEDML to each microcentrifuge tube containing a weighted clot. The lyophilized form of streptokinase, obtained from Incepta Pharmaceutical Ltd. in Dhaka, Bangladesh, was reconstituted using 2.5 mL of phosphate-buffered saline (PBS) and thoroughly mixed. To serve as a positive control, a volume of 100 µL of the aforementioned suspension was introduced into the microcentrifuge tube. Negative control was employed using a volume of 100 µL of distilled water. Following a 90minute incubation period at 37°C, an assessment was conducted to determine the presence of clot lysis within the tubes. Following incubation, the tubes were re-weighed in order to quantify the extent of weight alteration resulting from the disintegration of the clot. The percentage of clot lysis was determined through the measurement of weight variation prior to and post-clot lysis. The percentage of clot-lysis was calculated using the formula given below:

% of Clot lysis = 
$$\frac{A}{B} \times 100$$

Where, A and B represent the weight of released clot before and after treatment.

## 2.7 Cytotoxicity Test

#### 2.7.1 Brine shrimp lethality test

The brine shrimp lethality test [12] was employed as the standard bioassay to assess the cytotoxic properties of EEDML, a method commonly used for screening bioactive compounds. This study utilized Artemia salina as a zoological organism model. Initially, the procurement of brine shrimp eggs was carried out by acquiring them from a commercial establishment specializing in pet located in Dhaka. Bangladesh. supplies Following a 48-hour incubation period in a synthetic seawater medium consisting of a 3.8% sodium chloride solution, the brine shrimp successfully hatched and underwent development into larval shrimp, also known as nauplii. The methodology utilized for assessing the cytotoxicity of brine shrimp nauplii involved the application of Meyer's technique. In order to prepare the samples for testing, the EEDML was dissolved in a dimethyl sulfoxide solution with a maximum concentration of 50 µL per 5 mL. To attain concentrations of 0.98, 1.95, 3.91, 7.81, 15.625, 31.25, 62.5, 125, 250, 500µg/mL, a 3.8% NaCl solution was supplemented up to a volume of 5 mL. In this study, the positive control utilized as vincristine sulfate, which is a standard medication. Each test tube was populated with ten fully-grown shrimps. Following a 24-hour incubation period, the vials were subjected to visual inspection utilizing a magnifying glass to determine the number of viable nauplii. The LC<sub>50</sub> value was determined through the utilization of a logarithmic graph plotting the concentration levels against the corresponding mortality rates.

#### 2.8 Experimental Animals for *In-vivo* Pharmacological Investigation

A cohort of young (6-7 weeks) and healthy *Swiss albino* mice, with a weight range of 22-30g, was chosen as the experimental subjects.

The mice were procured from the Saver facility of Jahangirnagar University located in Dhaka, Bangladesh. Maintaining the existing state of affairs was deemed crucial. Typical variations in the atmosphere encompass a temperature range of approximately 79°F, a relative humidity level ranging from 55% to 65%, and a consistent 24-hour light/dark cycle. The specimens are subjected to consistent environmental conditions for a period of 9 days subsequent to their collection. In order to aid in the recovery of mice from water and food deprivation incurred during transportation and to facilitate their adjustment to

the laboratory setting, a diet consisting of appropriate nourishment and uncontaminated water was provided, in accordance with the guidelines prescribed by Jahangirnagar University. Following a 12-day period of recovery, the mice were deemed prepared for the commencement of the experiment.

#### 2.9 Analgesic Investigation

#### 2.9.1 Hot plate test

The experiment involved the utilization of a hot plate analgesia meter, specifically Eddy's Hot Plate using the method from previously described paper, to conduct the hot plate test [16]. The mice were assigned to four groups in a random manner, with each group consisting of five mice. The groups were as follows: the model group, the Diclofenac sodium group (5 mg/kg), the low-dose EEDML group (200 mg/kg), and the high-dose EEDML group (400 mg/kg). The nociceptive latency, which refers to the time it takes to respond to a painful stimulus, was measured by observing behaviors such as jumping or paw licking in mice placed on a heated surface at a constant temperature of 55 ± 0.1°C. After administering treatments, the latencies were measured and considered the baseline value. Only mice that exhibited initial pain responses within a time range of 5 to 20 seconds were chosen for inclusion in the experiment. The reaction time of each animal after receiving EEDML or Diclofenac sodium was measured at three-time intervals: 30, 60, and 120-minutes post-treatment. The designated time limit was established at 20 seconds.

The formula for the analgesic effectiveness of treatment was as follows:

Percent Analgesic Score = 
$$\frac{Ta-Tb}{Ta}$$
 X100

Time (in seconds) to react (before medication administration): Tb; Time (in seconds) to react (after drug administration): Ta.

#### 2.9.2 Acetic-acid induced writhing test

The experiment involved conducting the acetic acid-induced abdominal writhing test on mice in order to evaluate the analgesic effects of EEDML [17]. The mice were divided into five groups (n = 5) according to the hot plate test protocol. In this study, the mice were subjected to intraperitoneal administration of diclofenac sodium and EEDML one hour prior to the induction of writhing. The

writhing was induced by intraperitoneal injection of 0.6% acetic acid at a dosage of 10 mL per kg of body weight. Individual mice were placed in a spacious glass enclosure, and the frequency of writhing movements was recorded during a 10minute interval. The writhing was determined using the following formula:

% Of writhing = 
$$\frac{VC - VT}{VC} \times 100$$

Where, VT = number of writhing motions in extract-treated mice. VC = number of writhing motions in the control group of mice.

## 2.10 Statical Statistical Analysis

The bioassay readings were conducted twice, and the tabular data presented represent the mean value. The statistical analyses were performed utilizing the Microsoft Excel.

#### 3. RESULTS AND DISCUSSION

## 3.1 Anti-inflammatory Activity

Results from the present study showed that both MEPLL and EEDML exhibited significant effects when compared with conventional acetyl-salicylic acid, as shown in Table 1.

The anti-inflammatory effects of Diospyros malabarica were evaluated using the antialbumin denaturation technique. of egg Furthermore. studies have previous demonstrated that conventional nonsteroidal anti-inflammatory drugs (NSAIDs), including phenylbutazone and indomethacin, not only reduce the synthesis of endogenous prostaglandins by inhibiting the COX enzyme but also hinder protein denaturation [18]. Given the circumstances, the anti-denaturation assay offers a pragmatic approach to assess the presence of anti-inflammatory characteristics. The findings of the current study indicate that the extract possesses strong anti-inflammatory properties. A comparative analysis was conducted to evaluate

the effectiveness of *Diospyros malabarica* in regulating auto antigen synthesis and preventing protein denaturation, in comparison to the standard medication salicylic acid.

## 3.2 Anti-microbial Activity

The antibacterial and antifungal activity of various dosages of plant extract was evaluated gram-positive against and gram-negative bacteria, as well as four different types of fungi, by measuring the zone of growth inhibition in millimeters (mm). The findings indicate that there was a positive correlation between the concentration of plant extract and the size of the inhibitory zone, as observed in Table 2 and Table 3. The antibacterial activity exhibited a zone of inhibition ranging from 08 to 18 mm, while the zone of inhibition for fungi ranged from 06 to 14 mm. The results presented indicated that EEDML had the highest level of antibacterial activity against Escherichia coli and Bacillus cereus. The E. coli and B. cereus are known to cause diseases such as urinary tract infections, respiratory illness, pneumonia, gastrointestinal illness, and diarrhea.

The assessment of antimicrobial agents' efficacy necessitates the inclusion of antibacterial and antifungal tests. Antibacterial tests are conducted to assess the capacity of a substance to hinder the proliferation of bacteria, while antifungal tests are performed to evaluate the efficacy of a substance in suppressing the growth of fungi [19]. The agar diffusion method, also known as the Kirby-Bauer method, is widely used for antibacterial activity. Durina testina the experimental procedure, bacteria are cultivated on an agar plate, followed by the placement of discs containing the antimicrobial agent onto the agar's surface. The magnitude of the zone of inhibition surrounding each disc is indicative of the level of bacterial growth inhibition. The measurement of the zone of inhibition's diameter utilized determine the antimicrobial is to susceptibility of the bacteria. This measurement is then compared with a standard chart.

Table 1. Percentage	inhibition in	egg albumin	denaturation	of EEDML

Concentration	% Inhibition of acetyl salicylic acid	% Inhibition of extract EEDML
62.5	89.06	73.63
125	91.89	80.90
250	93.69	84.54
500	94.59	76.36
1000	98.19	85.45

Diameter of zone of inhibition (mm)						
Test organismsEEDMLEEDMLEEDMLCi(300 μg/disc)(500 μg/disc)(700 μg/disc)						
Gram Positive Bacteria						
Bacillus cereus	08	09	11	25		
Staphylococcus aureus	08	09	10	26		
Gram Negative Bacteria						
Escherichia coli	07	07	09	25		
Vibrio cholerae	11	08	08	27		
Klebsiella pneumonia	07	08	09	24		

Table 2. Inhibition zone of EEDML against different bacteria

Table 3. Inhibition zone of EEDML against different fungi

Diameter of zone of inhibition (mm)					
Test organisms	MELA (300 μg/disc)	MELA (500 μg/disc)	MELA (700 μg/disc)	Griseofulvin (50 µg/disk)	
Fungi					
Penicillium	07	10	15	19	
chrysogenum					
Aspergillus niger	08	09	11	20	
Yeast budding	9	11	15	21	
Mucor hiemalis	08	10	12	21	

Antifungal tests can be conducted using the agar diffusion method, which bears resemblance to the Kirby-Bauer method employed for bacterial tests. The agar diffusion method involves the cultivation of a fungal strain on an agar medium. Subsequently, discs containing the antifungal agent are positioned on the surface of the agar. The measurement of zone of inhibition surrounding each disc is utilized to assess the antifungal activity of the substance.

In conclusion, the evaluation of antimicrobial agents' effectiveness relies heavily on conducting antibacterial and antifungal tests. The agar diffusion and broth dilution methods are frequently employed for assessing the extent of bacterial and fungal inhibition or kill rate caused by various substances. These tests play a crucial role in the development and assessment of novel antimicrobial agents, as well as in the identification of appropriate agents for the management of bacterial and fungal infections.

## 3.3 Thrombolytic Activity

The efficacy of plant extracts in the dissolution of blood clots were assessed through an in vitro thrombolysis assay conducted under controlled laboratory conditions. In the event of bodily injury, the process of hemostasis is initiated, whereby blood cells and proteins aggregate to form a coagulum, thereby impeding the continuation of hemorrhaging. Nevertheless, an overabundance of clotting can lead to lifethreatening conditions such as myocardial infarctions and cerebrovascular accidents. Table 4 illustrates a notable disparity in the percentage of clot lysis between EEDML and conventional streptokinase.

## Table 4. Mean value of percent of clot lysis (N=10)

Sample	% of clot lysis
Negative control	7.296
Streptokinase	91.304
EEDML	85.37

Previous studies [12] have been conducted to investigate the thrombolytic potential of supplements, herbs, and natural food sources in the treatment of coronary events and strokes. The thrombolytic efficacy of EEDML was observed to be rapid, with a value of 85.37%, as compared with the standard value of 91,304% (Table 4). The observed value is derived from the fact that EEDML reduces the coagulation of human blood in vitro, thereby suggesting its potential as a cardioprotective agent. The EEDML exhibited considerable significance, which could have significant implications for cardiovascular health. Consequently, it might facilitate the development of novel thrombolytic agents derived from Diospyros malabarica [20].

## **3.4 Cytotoxic Activity**

The brine shrimp lethality assay is a commonly employed bioassay in the field of plant extract evaluation, serving as an initial screening method to assess the biological activity of such extracts. The purpose of this test is to evaluate the toxicity of compounds and assess the cytotoxic properties of plant extracts. The initial stage of plant screening involves the assessment of cytotoxicity, which provides valuable insights into the potential antitumor and anticancer properties of the plant extract. The cytotoxic effects of EEDML on the growth of brine shrimp were investigated.

The brine shrimp assay represents a costeffective and straightforward approach to assessing the cytotoxic characteristics of plant extracts. The method employed in this study was used to assess the cytotoxic activity of EEDML. The findings obtained from this analysis have been succinctly presented in Table 4. The LC<sub>50</sub> values for EEDML and the standard drug Vincristine Sulphate were determined to be 1.147 µg/mL and 0.608 µg/mL, respectively, as shown in Fig. 1. Furthermore, various concentrations of the test solution exhibited varying levels of mortality in Artemia salina. The LC<sub>50</sub> values exhibited a range of 1.95 µg/mL (statistically significant) to 500 µg/mL (highly statistically significant), indicating a clear relationship between concentration and  $LC_{50}$ . The highest mortality rate was observed at a concentration of

500 µg/mL, whereas the lowest mortality rate was observed at a concentration of 1.147 µg/mL. It can be posited that there exists a positive correlation between the concentration of the test samples and the percentage of mortality, whereby an increase in concentration leads to a corresponding increase in mortality, and conversely, a decrease in concentration results in a decrease in mortality. In comparison to the standard vincristine sulfate concentration of the EEDML 0.608 µg/mL, demonstrates significant cytotoxicity towards brine shrimp nauplii, as evidenced by its LC<sub>50</sub> value of 1.147 µg/mL. The cytotoxicity of EEDML exhibits notable differences when compared to the standard vincristine sulfate. This finding warrants further investigation to explore its potential as a compound for antitumor and pesticide applications [21].

## 3.5 *In-vivo* Analgesic Activity

#### 3.5.1 Hot plate test

In this experiment, the EEDML exhibited a significant analgesic activity that varied in potency and dosage dependence when compared to the standard Morphine Sulphate. The analgesic activity of the extract was determined to be slightly more than half of that exhibited by the standard. The average time tolerance values for the extract and standard are provided in Table 6.

Sample name	Concentrations	Mortality %	LC <sub>50</sub> value
	7.81	40	
	15.625	40	
	31.25	50	20.57
Vincristine Sulphate	62.5	60	
	125	100	
	250	100	
	500	100	
	0.98	10	
	1.95	30	
	3.91	40	
	7.81	40	1.147
EEDML	15.625	50	
	31.25	70	
	62.5	70	
	125	80	
	250	100	
	500	100	

Table 5	. Brine	shrimp	assay	(Mortality	%, LC <sub>50</sub>	value)
---------	---------	--------	-------	------------	---------------------	--------





#### Fig. 1. Cytotoxic activity of EEDML on brine shrimp nauplii

	Reaction time at different	time interva	ls (in sec)		
Group	Average wt. of mice (g)	30 min	60 min	90 min	120 min
Control		6.4	7.6	6.0	5.4

8.6

9.6

33.39

9.4

12.6

	to 26	12.2	13.2	0	0
Table 7. Analgesic activity of EEDML on mice by writhing test					
Administered substance	Dose	% Writ	hing	% Of	inhibition
Control	10mL/kg	100		0.00	
Diclofenac sodium	10mg/kg	20.40		79.61	
EEDML	200mg/kg	53.80		46.2	

400mg/kg

The hot plate test is a commonly employed experimental paradigm utilized for the identification of compounds that demonstrate centrally acting analgesic effects. Multiple animal species have demonstrated that the hot plate method exhibits selectivity towards opioid-related compounds [22]. There exists speculation regarding the potential role of these molecules in mediating the narcotic analgesic, antiinflammatory, and anti-diabetic effects. The findings from the experiment indicate that there is a correlation between the dosage of EEDML and the amount of time it takes for thermal pain to be perceived. This suggests that the analgesic effects of EEDML may be attributed to a similar underlying mechanism.

Morphine (5mg/kg)

EEDML (200mg/kg)

EEDML

## 3.5.2 Results of acetic-acid induced writhing test

11.0

14.8

66.61

6.6

0

In the acetic acid-induced writhing test, the methanolic extracts of *Diospyros malabarica* leaf exhibited inhibitory effects of 66.61% and 46.2% respectively. The standard drug, Diclofenac-Na, demonstrated an inhibition rate of 80.36%. In comparison to established standards, the findings exhibited a noteworthy level of analgesic efficacy that is contingent upon the dosage administered.

The extract of Diospyros malabarica demonstrated a significant and dose-dependent reduction in abdominal writhing during an acetic

acid-induced abdominal constriction test. The efficacy of the test in assessing the effectiveness of moderate analgesic non-steroidal antiinflammatory drugs (NSAIDs) is attributed to the indirect mechanism of acetic acid. Acetic acid is believed to induce the release of prostaglandins and lipo-oxygenase products into the peritoneum, thereby activating nociceptive neurons that are responsive to NSAIDs. The findings from the experiment involving acetic provide writhina acid-induced compelling evidence that the observed effect is partially associated with the inhibition of lipo-oxygenase and/or cyclooxygenase in the peripheral tissues. This inhibition leads to a reduction in prostaglandin synthesis, which in turn disrupts the transduction mechanism in primary afferent nociceptors.

## 4. CONCLUSION

The methanolic extract of Diospyros malabarica's leaf has been found to possess antiantimicrobial. thrombolvtic. inflammatory, cytotoxic, and analgesic properties, leading to the inference that it may have potential therapeutic applications in these areas. The dose-dependent anti-inflammatory effect of this plant observed in the present study was notably significant when compared with the standard. The leaf extract demonstrated a minimal level of antimicrobial activity while the thrombolytic effect exhibited was found to be highly significant. The findings from the bioassay conducted on the lethality of brine shrimp indicated a significant level of cytotoxicity. Both the writhing test and the hot plate test provide evidence of the analgesic effects exhibited by this plant. Furthermore, it has been established that administering a higher dosage yields greater efficacy compared to a lower dose.

## ACKNOWLEDGEMENT

The authors would like to acknowledge the Laboratory of Pharmacology at Stamford University Bangladesh and the Laboratories of Bangladesh Council of Scientific and Industrial Research for their valuable support, provision of resources, and provision of necessary infrastructural facilities that were essential for the successful execution of this research study.

## CONSENT

It is not applicable.

## ETHICAL APPROVAL

The authors declare that all experiments were carried out in adherence to ethical guidelines and underwent review by a suitable ethics committee.

## **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

## REFERENCES

- Cragg GM, Newman DJ. Natural products: A continuing source of novel drug leads. Biochimica et Biophysica Acta - General Subjects. 2013;1830(6):3670–3695. DOI:https://doi.org/10.1016/j.bbagen.2013. 02.008
- Kilmer PD. Review article: Review article. Journalism. 2010;11(3):369–373. DOI:https://doi.org/10.1177/146144481036 5020
- White NJ. Qinghaosu (artemisinin): The price of success. Science. 2008; 320(5874):330–334. DOI:https://doi.org/10.1126/science.11551 65
- Ramawat KG, Dass S, Mathur M. Herbal drugs: Ethnomedicine to modern medicine; 2009.
- Dawid-Pać R. Medicinal plants used in treatment of inflammatory skin diseases. Postepy Dermatologii i Alergologii. 2013; 30(3):170–177. DOI:https://doi.org/10.5114/pdia.2013.356 20
- Mhalhel K, Sicari M, Pansera L, Chen J, Levanti M, Diotel N, Rastegar S, Germanà, A, Montalbano G. Zebrafish: A model deciphering the impact of flavonoids on neurodegenerative disorders. Cells. 2023; 12(2).

DOI:https://doi.org/10.3390/cells12020252

- Tu Y. Artemisinin—A gift from traditional Chinese medicine to the World (Nobel Lecture). Angewandte Chemie -International Edition. 2016;55(35):10210– 10226. DOI:https://doi.org/10.1002/anie.20160196 7
- Tripathi MK, Singh P, Sharma S, Singh TP, Ethayathulla AS, Kaur P. Identification of bioactive molecule from *Withania somnifera* (Ashwagandha) as SARS-CoV-2 main protease inhibitor. Journal of

Biomolecular Structure and Dynamics. 2020;0(0):1–14. DOI:https://doi.org/10.1080/07391102.202 0.1790425

- Polash SA, Hamza A, Hossain MM, Tushar MH, Takikawa M, Shubhra RD, Saiara N, Saha T, Takeoka S, Sarker SR. *Diospyros malabarica* fruit extract derived silver nanoparticles: A biocompatible antibacterial agent. Frontiers in Nanotechnology. 2022 4 May. DOI:https://doi.org/10.3389/fnano.2022.88 8444
- Moniruzzaman M, Kuddus MR, Chowdhury AS, Rashid MA. Antioxidant, antimicrobial, anti-diarrheal and analgesic activities of *Diospyros malabarica* (Desr.) Kostel. Bangladesh Pharmaceutical Journal. 2019; 22(1):27–33. DOI:https://doi.org/10.3329/bpj.v22i1.4002

DOI:https://doi.org/10.3329/bpj.v22i1.4002

- Kavatagimath SA, Jalalpure SS. Screening of ethanolic extract of diospyros malabarica desr. Bark for anti-diabetic and antioxidant potential. Indian Journal of Pharmaceutical Education and Research. 2016;50(1):179–189. DOI:https://doi.org/10.5530/ijper.50.1.22
- Shaira HA, Shomudro HK, Chowdhury SA. In-vitro and In-vivo Pharmacological evaluation of Persicaria lapathifolia available in Bangladesh. Journal of Scientific Research and Reports. 2023; 29(3):12–26. DOI:https://doi.org/10.9734/jsrr/2023/v29i3 1733
- Arif Ullah HM, Sayera Zaman, Fatematuj Juhara, Lucky Akter, Syed Mohammed Tareq, Emranul Haque Masum RB. Antiinflammatory activity of ethanolic extract of *Curcuma zedoaria* rhizome. BMC Complementary and Alternative Medicine. 2014;14:1–12
- Somchit MN, Reezal I, Elysha Nur I, Mutalib AR. *In vitro* antimicrobial activity of ethanol and water extracts of Cassia alata. Journal of Ethnopharmacology. 2003; 84(1):1–4. DOI:https://doi.org/10.1016/S0378-8741(02)00146-0

- Antioxidant I, Mill S. Neuropharmacological activities of bioactive metabolites of. 2023; 35(7):29–39. DOI:https://doi.org/10.9734/JPRI/2023/v35i 77338
- Nguyen T, Chen X, Chai J, Li R, Han X, Chen X, Liu S, Chen M, Xu X. Antipyretic, anti-inflammatory and analgesic activities of Periplaneta americana extract and underlying mechanisms. Biomedicine and Pharmacotherapy. (September 2019). 2020;123:109753. https://doi.org/10.1016/j.biopha.2019.1097 53
- Wang J. wen, Chen S. shan, Zhang Y. meng, Guan J Su GY, Ding M, Li W, Zhao YQ. Anti-inflammatory and analgesic activity based on polymorphism of cedrol in mice. Environmental Toxicology and Pharmacology. (September 2018). 2019; 68:13–18. DOI:https://doi.org/10.1016/j.etap.2019.02

DOI:https://doi.org/10.1016/j.etap.2019.02. 005

- 18. Yamamoto H, Matsumoto K, Araki E, Miyamoto aspects Ε. New of neurotransmitter release and exocytosis: Involvement Ca2+/calmodulinof dependent phosphorylation of synapsin I in insulin exocytosis. Journal of Pharmacological Sciences. 2003;93(1): 30-34.
- Herbal drugs: Ethnomedicine to modern medicine; June 2014. DOI:https://doi.org/10.1007/978-3-540-79116-4
- 20. Ratnasooriya WD, Fernando TSP, Madubashini PP. *In vitro* thrombolytic activity of Sri Lankan black tea, Camellia sinensis (L.) O. Kuntze. 2008;36(2):36–38.
- 21. Suffredini IB, Paciencia MLB, Nepomuceno DC, Younes RN, Varella AD. Antibacterial and cytotoxic activity of Brazilian plant extracts - Clusiaceae. May 2006;101:287–290.
- 22. Menyiy E, Balahbib A, El-shazly M, Bakri Y. na r f. Journal of Pharmaceutical Analysis; 2021. DOI:https://doi.org/10.1016/j.jpha.2021.07. 004

© 2023 Chowdhury et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/104196