



Studies on the Root Architecture with Nodulation of the Chickpea (*Cicer arietinum* L.) as Influence by Different Moisture Management Practices along with Seed Inoculation and Level of Zinc

Kishan Kumar ^{a*}, Ram Pyare ^a, Ram Niwas ^a, Kamal Tiwari ^a, Ravindra Sachan ^b, Ravikesh Kumar Pal ^c, Vinay Kumar Patel ^d and Abhishek Raj Ranjan ^d

^a Department of Agronomy, Chandra Shekhar Azad University of Agriculture and Technology, Kanpur (U.P.), 208002, India.

^b Department of Soil Science and Agricultural Chemistry, Chandra Shekhar Azad University of Agriculture and Technology, Kanpur (U.P.), 208002, India.

^c Department of Agronomy, B.A.U. Sabaur, Bhagalpur (Bihar), 813210, India.

^d Department of Agronomy, Acharya Narendra Dev University of Agriculture and Technology, Kumarganj (Ayodhya), 224229, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJECC/2022/v12i1131282

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/92373>

Original Research Article

Received 15 July 2022

Accepted 26 September 2022

Published 01 October 2022

ABSTRACT

The current field experiment was carried out during *Rabi* season of 2020-21 and 2021-22 at the Student's Instructional Farm of Chandra Shekhar Azad University of Agriculture and Technology, Kanpur, Uttar Pradesh to assess the Studies on the root architecture with nodulation of the chickpea (*Cicer arietinum* L.) as influence by different moisture management practices along with seed inoculation and level of zinc. The experiment was laid out in split-split plot design with 27 treatment combination comprising three moisture conservation practices namely, flat bed with 2.5 t/ha crop residue, narrow bed and furrow with 2.5 t/ha crop residue, broad bed and furrow with 2.5 t/ha crop residue in main plot and three seed inoculation (control, *rhizobium* and *PSB*) in sub-plots and three zinc level (control, 2.5 kg zinc/ha and 5.0 kg zinc/ha) in sub- sub plot with three

replication. Results showed that among the different moisture management practices, used of broad bed and furrow with 2.5 t/ha crop residue were significantly enhanced root architecture, nodulation and grain yield, over the flat bed with 2.5 t/ha crop residue, respectively. Among the different Biofertilizers treatments seed inoculation with *Rhizobium* had significantly improved the root architecture, nodulation and yield, over control. Application of increasing levels of zinc up to 5.0 kg Zn/ha has significant influences on root architecture, nodulation and yield, over control. The combined application broad bed and furrow with 2.5 t/ha crop residue with *Rhizobium* and 5.0 kg/ha zinc resulted in significantly higher root architecture, nodulation and seed yield of chickpea during both the years of experimentation.

Keywords: Broad bed and furrow; chickpea; narrow bed and furrow; nodule; rhizobium and zinc.

1. INTRODUCTION

Chickpea (*Cicer arietinum* L.) occupies prominent position among the various pulse crop grown in India. India ranks first in the world in respect of production as well as acreage and produces 11.23 million tons chickpea grains from 10.56 million hectare area with an average productivity of 1063 Kg ha⁻¹ during 2017-18. India contributes 71 per cent of chickpea production of the world [1]. It is commonly used for human consumption as well as for feeding animals. Chickpea is considered to have medicinal effects and it is used for blood purification. Chickpea is free from various anti-nutritional factors and has high protein (18-22%), total carbohydrates (52-70%) and fat (4-10%). It is also a rich source of minerals (calcium, phosphorus, iron, niacin) and vitamins. Being a leguminous crop it has unique property of maintaining and restoring soil fertility through biological nitrogen fixation and also improve soil organic matter by addition of ample amount of residue due to shedding of their leaves at maturity.

Chickpea mostly grown on stored or residual soil moisture after harvest of *kharif* crops faces moisture stress throughout the life cycle. Nitrogen fixation by leguminous plants is reduced by moisture stress due to reduction in leghaemoglobin in nodules, specific nodule activity and number of nodules. The risk factor can be minimized through *in situ* moisture conservation, adoption of suitable crops and their varieties [2]. *In-situ* application of crop residues and division of field into beds and furrows could be used as low-cost input technology, which helps to conserve more rainwater in soil by minimizing runoff of water from soil surface under water scarcity situations [3]. Application of crop residue on soil surface as a mulch reduces the loss of water through evaporation and moderate the soil profile temperature [4].

Seed inoculation with *Rhizobium* increase the nodulation through better root development and improves nutrient availability which is beneficial in improving the grain yield. Phosphorus solubilizing bacteria is a cheapest source of phosphorus availability particularly in legume crops, it possess the ability to bring sparingly insoluble organic and inorganic phosphate into soluble forms by secreting organic acid [5-7]. Use of biofertilizer such as *rhizobium* can reduce the need for chemical fertilizer and decrease adverse environmental effects. Biofertilizers provide an economically judicious, attractive and ecologically sound means of fertilization [8]. Chickpea Inoculated with *Rhizobium* significantly increased the nodulation and its dry weight, root length, root dry weight and grain yield [9]. Rhizobium and phosphate solubilizing bacteria (PSB) assume a great importance on account of their vital role in N₂ fixation and P solubilization [10-12].

Zinc has an important metabolic role in plant growth and development and therefore, called an essential trace element or micronutrient [13,14]. Zinc is involved in various host plant metabolic processes, nodule growth and N₂ fixation process. Zn nutrient is receiving substantial attention as application of zinc in many legumes has also been found to increase root growth, nodulation and yield [15]. In addition to having an important role in activating plants enzymatic systems. Zinc is essential for the synthesis of chlorophyll and carbohydrates. This element plays an important role in increasing plant resistance to fungal disease and expanding plant roots [16]. Many area of the world suitable for chickpea have widespread zinc deficiency. Zinc deficiency affect plant water relation, induce stomata closure and decreases transpiration in plant. Chickpea is generally considered sensitive to zinc deficiency compared to various crop species. Zinc deficiency reduces not only the grain yield but also nutritional quality of the grain.

2. MATERIALS AND METHODS

The experiment was conducted at “Students Instructional Farm” of Department of Agronomy, Chandra Shekhar Azad University of Agriculture & Technology, Kanpur-208002 (U.P.) during the *Rabi* season of 2020-21 and 2021-22. The experiment was laid out in spit-split plot design with three replication. The experiment was conducted in with 27 treatment combination comprising three moisture conservation practices namely, flat bed with 2.5 t/ha crop residue, narrow bed and furrow with 2.5 t/ha crop residue, broad bed and furrow with 2.5 t/ha crop residue in main plot and three seed inoculation (control, *rhizobium* and PSB) in sub-plots and three zinc level (control, 2.5 kg zinc/ha and 5.0 kg zinc/ha) in sub- sub plot. The chickpea variety RVG 202 was used for field experiment during both the year. The crop was fertilized as per the treatment. The recommended dose of nitrogen, phosphorus and potassium @ Recommended dose of fertilizers were applied to the crops during both the years in all plots. 20 kg N, 40 kg P₂O₅ and 40 kg K₂O/ha were applied in all the plots as basal dose at the time of sowing. Urea, DAP, Murate of potash were used as the source of nitrogen, phosphorus and potassium. After field preparation and before sowing of crop, the narrow beds of 70 cm wide with furrows of 30 cm width and broad beds of 90 cm wide with furrow of 30 cm width were prepared manually in respective plots. Paddy straw residue was applied in chickpea crop as per treatments just after sowing as moisture management treatments during both the years of study. Zinc was applied as per treatments through zinc sulphate (ZnSO₄.7H₂O) containing 21% Zn and 10% S at the time of sowing as basal dose. seeds of chickpea are inoculated with *Rhizobium* and PSB as per treatments one day before sowing treated seeds are spread in shades for 8-10 hours then after used for sowing.

3. RESULT AND DISCUSSION

3.1 Root Architecture and Root Length

It is visualized from the data given in Table 1, higher values of root length at nodulation and flowering stage of chickpea was observed under broad bed and furrow with 2.5t/ha crop residue in both the years and in pooled data. sowing of chickpea under broad bed and furrow with 2.5t/ha crop residue resulted into significantly higher root length (7.12 and 28.32

cm) at nodulation and flowering stage as compared to other moisture conservation practices in pooled data study with percent increment 13.37, 34.98 over flat bed with 2.5 t/ha crop residue at nodulation and flowering stage, respectively. The results of present investigation are also in agreement with the findings of Rathore et al. [2], Mishra et al. [17], Lal et al. [18], Kumar et al. [19], Chavan et al. [20] and Gupta et al. [21].

The data presented in Table 1 clearly indicate that response of seed inoculation to root length show significant variation in length of root was increased in rhizobium as compare to control and PSB. The percentage increment over control of 3.35, 9.41 and PSB of 1.49, 4.21 at nodulation and flowering stage in pooled analyzed data of experimentation, respectively. These results also confirms of the findings of Shahzad et al. [9], Chandra and Pareek [22], Gyandev et al. [23], Khaitov et al. [24], Chauhan and Singh [25], Singh et al. [26], Katiyar et al. [27], Benjelloun et al. [28] and Yadav et al. [29].

Application of increasing levels of zinc up to 5.0 kg Zn/ha did not influences the root length at nodulation stage whereas it significantly influenced at flowering stage in 2020-21 and 2021-22 (Table 1). However, direct application of chickpea with 5.0 Zn/ha enhanced the root length by 0.59, 1.38 and 1.05, 2.42 percent at nodulation and flowering stage, respectively, over 2.5 kg/ha zinc and control in pooled data of experimentation. Similar result was reported by Gupta et al. [30], Yadav et al. [31] and Singh and Bhati [32].

3.2 Dry Weight of Root

Data revealed that higher values of root dry weight at nodulation and flowering stage of chickpea was observed under broad bed and furrow with 2.5t/ha crop residue (Table 1) in 2020-21 and 20221-22 as well as in pooled. Sowing of chickpea under broad bed and furrow with 2.5t/ha crop residue resulted into significantly higher root dry weight (0.924 and 2.48 g) at nodulation and flowering stage as compared to other moisture conservation practices with percent improvement 62.67 and 16.98 over flat bed with 2.5 t/ha crop residue at nodulation and flowering stage. These results also confirms of the findings of Rathore et al. [2], Mishra et al. [17], Lal et al. [18], Kumar et al. [19], Chavan et al. [20] and Gupta et al. [21].

The data clearly indicate that response of seed inoculation to dry weight of root show significant variation, dry weight of root was increased in *rhizobium* as compare to control and PSB. The percentage increment over control of 16.76, 4.91 and PSB of 8.16, 2.62 at nodulation and flowering stage in both the years of studies, respectively. Similar result was reported by Verma et al. [33], Verma et al. [34], Abisha and Singh [35] and Yadav et al. [29].

Application of increasing level of zinc up to 5.0 kg Zn/ha has significant influences on dry weight of root at nodulation while it became at par at flowering in pooled analyzed data (Table 1). However, direct application of chickpea with 5.0 Zn/ha enhanced the root dry weight by 2.52, 0.86 and 6.34, 2.20 percent at nodulation and flowering stage, respectively, over 2.5 kg/ha zinc and control in both the years of pooled data, respectively. The results of present investigation are also in agreement with the findings of Gupta et al. [30], Yadav et al. [31] and Singh and Bhati [32].

3.3 Number of Nodule

Data presented in Table 2 revealed that sowing of chickpea under broad bed and furrow with 2.5 t/ha crop residue proved significantly superior over narrow bed and furrow with 2.5 t/ha crop residue and flat bed with 2.5 t/ha crop residue in both the years of study. The significantly higher number of root nodules per plant (17.39, 17.48) were observed under broad bed and furrow with 2.5 t/ha crop residue during 2020-21 and 2021-22 over rest of the treatment with The percentage increment over flat bed with 2.5 t/ha crop residue of 24.30 in study of pooled data. The consequences of the current investigation are additionally in concurrence with the investigation of Mishra et al. [17], Lal et al. [18], Kumar et al. [19], Chavan et al. [20] and Gupta et al. [21].

The data clearly indicate that response of number of nodule per plant of chickpea to used seed inoculation was increased in *rhizobium* compare to control and PSB during both year of study (Table 2). The significantly higher number of root nodules per plant (16.23, 16.34) were observed under *rhizobium*. The results of present investigation are also in agreement with the findings of Khaitov et al. [24], Chauhan and Singh [25], Singh et al. [26], Verma et al. [33],

Katiyar et al. [27], Verma et al. [34] and Benjelloun et al. [28].

Number of root nodules per plant were also affected significantly due to zinc level treatments directly applied to chickpea (Table 2). Application of 5.0 kg Zn/ha to chickpea being statistically at par with 2.5 kg Zn/ha, recorded significantly higher number (15.96 and 16.05) over control during both the years of experimentation with percent increment of 1.20 in pooled study. These results also confirms of the findings of Singh and Bhati [32], Chaudhary et al. [36], Thenua et al. [37] and Yadav et al. [38].

3.4 Nodule Dry Weight

A perusal of data presented in Table 2 revealed that sowing of chickpea under broad bed and furrow with 2.5 t/ha crop residue produces significantly higher dry weight of root nodules per plant (208.24, 214.60 mg) during 2020-21 and 2021-22. The percent improvement in broad bed and furrow with 2.5 t/ha crop residue 13.34 over flat bed with 2.5 t/ha crop residue on pooled basis. These results also confirms of the findings of Mishra et al. [17], Lal et al. [18], Kumar et al. [19], Chavan et al. [20] and Gupta et al. [21].

The data clearly indicate that response of dry weight of root nodule to used seed inoculation was increased in *rhizobium* compare to control and PSB during both year of study. *Rhizobium* treated seed show significantly higher dry weight of root nodules per plant (199.08 and 206.02 mg) with percent improvement over control 3.75, 3.85 and PSB 1.77, 1.98 during both the years of experimentation, respectively (Table 2). The consequences of the current investigation are additionally in concurrence with the investigation of Khaitov et al. [24], Chauhan and Singh [25], Singh et al. [26], Verma et al. [33], Katiyar et al. [27], Verma et al. [34] and Benjelloun et al. [28].

Dry weight of root nodules per plant were also affected significantly due to zinc level treatments directly applied to chickpea. Application of 5.0 kg Zn/ha to chickpea being statistically at par with 2.5 kg Zn/ha and control in first years while in second year it was only at par with 2.5 kg Zn/ha, recorded significantly higher dry weight of root nodules per plant (196.80 and 204.43 mg) over control during both the years of experimentation, respectively (Table 2). The results of present investigation are also in agreement with the findings of Singh and Bhati [32], Chaudhary et al. [36], Thenua et al. [37] and Yadav et al. [38].

Table 1. Root length and root dry weight as influenced by Moisture conservation practices, Seed inoculation and Zinc level

Treatments	Root length (cm)						Dry weight of root g/plant					
	Nodulation stage			Flowering stage			Nodulation stage			Flowering stage		
	2020-21	2021-22	Pooled	2020-21	2021-22	Pooled	2020-21	2021-22	Pooled	2020-21	2021-22	Pooled
A. Moisture conservation practices												
Flat bed + 2.5 t/ha crop residue	5.82	6.74	6.28	20.51	21.44	20.98	0.559	0.577	0.568	2.11	2.13	2.12
NBF + 2.5 t/ha crop residue	6.13	7.11	6.62	24.07	25.00	24.53	0.744	0.766	0.755	2.28	2.30	2.29
BBF + 2.5 t/ha crop residue	6.62	7.60	7.12	27.86	28.77	28.32	0.917	0.932	0.924	2.46	2.49	2.48
S.Em. \pm	0.023	0.030	0.013	0.068	0.094	0.089	0.004	0.003	0.004	0.010	0.006	0.006
CD at 5%	0.089	0.118	0.049	0.265	0.367	0.347	0.017	0.010	0.014	0.037	0.023	0.023
B. Seed inoculation												
Control	6.08	7.03	6.56	23.03	23.93	23.48	0.686	0.699	0.692	2.23	2.26	2.24
Rhizobium	6.30	7.26	6.78	25.22	26.16	25.69	0.797	0.820	0.808	2.34	2.36	2.35
PSB	6.20	7.16	6.68	24.17	25.13	24.65	0.738	0.756	0.747	2.28	2.31	2.29
S.Em. \pm	0.023	0.033	0.016	0.064	0.080	0.075	0.003	0.003	0.002	0.011	0.008	0.008
CD at 5%	0.069	0.100	0.051	0.198	0.245	0.230	0.010	0.009	0.007	0.035	0.023	0.025
C. Zinc level												
Control	6.16	7.11	6.64	23.87	24.79	24.33	0.719	0.731	0.725	2.26	2.28	2.27
2.5 kg Zn/ha	6.19	7.15	6.67	24.11	25.04	24.58	0.742	0.761	0.752	2.29	2.31	2.30
5.0 kg Zn/ha	6.22	7.19	6.71	24.45	25.38	24.92	0.759	0.782	0.771	2.30	2.33	2.32
S.Em. \pm	0.034	0.038	0.026	0.085	0.091	0.077	0.003	0.004	0.004	0.012	0.009	0.008
CD at 5%	NS	NS	NS	0.243	0.260	0.220	0.010	0.012	0.010	0.033	0.027	0.024

Table 2. No. of nodule/plant, Nodule dry weight and grain yield as influenced by Moisture conservation practices, Seed inoculation and Zinc level

Treatments	Number of nodule/plant			Nodule dry weight(mg/plant)			Grain yield (q/ha)		
	2020-21	2021-22	Pooled	2020-21	2021-22	Pooled	2020-21	2021-22	Pooled
A. Moisture conservation practices									
Flat bed + 2.5 t/ha crop residue	13.98	14.07	14.03	183.27	189.80	186.53	13.61	13.89	13.75
NBF + 2.5 t/ha crop residue	16.24	16.31	16.28	195.04	202.01	198.53	15.12	15.71	15.42
BBF + 2.5 t/ha crop residue	17.39	17.48	17.44	208.24	214.60	211.42	16.75	17.53	17.14
S.Em. \pm	0.064	0.097	0.084	0.731	0.858	0.365	0.087	0.047	0.075
CD at 5%	0.251	0.379	0.329	2.856	3.348	1.427	0.338	0.184	0.291
B. Seed inoculation									
Control	15.48	15.53	15.50	191.87	198.37	195.12	14.62	15.15	14.89
Rhizobium	16.23	16.34	16.29	199.08	206.02	202.55	15.74	16.34	16.04
PSB	15.91	16.00	15.95	195.61	202.02	198.82	15.11	15.65	15.38
S.Em. \pm	0.076	0.088	0.072	0.713	0.920	0.497	0.064	0.059	0.047
CD at 5%	0.235	0.271	0.221	2.198	2.834	1.530	0.198	0.181	0.146
C. Zinc level									
Control	15.77	15.85	15.81	194.42	199.98	197.20	14.63	15.15	14.89
2.5 kg Zn/ha	15.89	15.97	15.93	195.33	202.00	198.67	15.18	15.73	15.45
5.0 kg Zn/ha	15.96	16.05	16.00	196.80	204.43	200.62	15.68	16.27	15.97
S.Em. \pm	0.053	0.054	0.052	0.989	1.083	0.784	0.070	0.082	0.074
CD at 5%	0.153	0.154	0.153	2.998	3.107	2.250	0.200	0.234	0.212

3.5 Grain Yield

The result revealed that grain yield was significantly increased at broad bed and furrow with 2.5 t/ha crop residue which was more than flat bed with 2.5 t/ha crop residue and narrow bed and furrow with 2.5 t/ha crop residue in first year and second year Table 2 with percentage increment over flat bed with 2.5 t/ha crop residue of 24.65 and narrow bed and furrow with 2.5 t/ha crop residue of 11.15 in pooled analyzed data of experimentation, respectively. The consequences of the current investigation are additionally in concurrence with the investigation of Kumar et al. [39], Chavan et al. [20] and Gupta et al. [21].

The data clearly indicate that response of grain yield to used seed inoculation was increased in *rhizobium* compared to control and PSB during both year of study Table 2. The percentage increment over control of 7.72 and PSB of 4.29 in pooled analyzed data of experimentation, respectively. The results of present investigation are also in agreement with the findings of Chauhan and Singh [25], Singh et al. [40], Singh and Singh [41], Singh et al. [42], Verma et al. [33], Verma et al. [34], Katiyar et al. [27], Benjelloun et al. [28] and Yadav et al. [29].

It is clear from the Table 2 that zinc application exerted a positive effect on grain yield where the significantly response noted up to 5.0 kg Zinc/ha in both the years with percentage increment over control of 7.25 in pooled analyzed data. These results also confirms of the findings of Singh et al. [43], Shivay et al. [44], Parmar et al. [45] and Yadav et al. [38].

4. CONCLUSION

Based on the above result, it can be concluded that the broad bed and furrow with 2.5 t/ha crop residue is superior over the remaining moisture management practices with use of seed inoculation of *rhizobium* and dose of 5.0 kg zinc/ha in respect to root architecture, nodulation, and seed yield. Thus broad bed and furrow with 2.5 t/ha crop residue and *rhizobium* with 5.0 kg zinc/ha may be recommended to realize higher root architecture, nodulation and grain yields of chickpea.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Anonymous. DES, Ministry of Agri. &FW (DAC&FW) Govt. of India; 2020.
2. Rathore RS, Singh RP, Nawange DD. Effect of land configuration, seed rates and fertilizer doses on growth and yield of black gram [*Vigna mungo* L.]. Legume Research. 2010;33(4):274-278.
3. Singh G, Sekhon HS, Kaur H. Effect of farmyard manure, vermicompost and chemical nutrients on growth and yield of chickpea (*Cicer arietinum* L.). International Journal of Agricultural Research. 2012; 7(2):93-99.
4. Ram H, Singh Y, Saini KS, Kler DS, Timsina J, Humphreys EJ. Agronomic and economic evaluation of permanent raised beds, no tillage and straw mulching for an irrigated maize-wheat system in northwest India. Experimental Agriculture. 2012; 48(1):21-38.
5. Pal V, Singh G, Dhaliwal SS. Agronomic biofortification of chickpea with zinc and iron through application of zinc and urea. Communication in Soil Science and Plant Analysis. 2019;50(15):1864-1877.
6. Ramesh T, Rathika S, Nagarajan G, Shanmugapriya P. Land configuration and nitrogen management for enhancing the crop productivity: A review; 2020.
7. Rudresh DL, Shivaprakasha MK, Prasad RD. Effect of combined application of *Rhizobium*, phosphate solubilizing bacterium and *Trichoderma* spp. on growth, nutrient uptake and yield of chickpea (*Cicer aritenium* L.). App Soil Ecol. 2005;28:139-46.
8. Patel PS, Ram RB, Jayprakash Meena ML. Effect of biofertilizers on growth and yield attributes of Pea (*Pisum sativum* L.) Trends in Biosciences. 2013;6(2):174-76.
9. Shahzad SM, Khalid A, Arif MS, Riaz M, Ashraf M, Iqbal Z, Yasmeen T. Co-inoculation integrated with P-enriched compost improved nodulation and growth of Chickpea (*Cicer arietinum* L.) under irrigated and rainfed farming systems. Biology and Fertility of Soils. 2014;50(1):1-12.
10. Jyothi CN, Ravichandra K, Babu KS. Effect of foliar supplementation of nitrogen and zinc on soybean (*Glycine max* L.) yield, quality and nutrient uptake. Indian Journal of Dryland Agricultural Research and Development. 2013;28(2):46-48.

11. Mishra JP, Praharaj CS, Singh KK, Kumar N. Impact of conservation practices on crop water use and productivity in chickpea under middle Indo-Gangetic plains. *Journal of Food Legumes*. 2012a;25(1):41-44.
12. Paliwal DK, Kushwaha HS, Thakur HS. Performance of soybean (*Glycine max*)-wheat (*Triticum aestivum*) cropping system under land configuration, mulching and nutrient management. *Indian Journal of Agronomy*. 2011;56(4):334-339.
13. Akhtar M, Siddiqui Z. Effects of phosphate solubilizing microorganisms and *Rhizobium* sp. on the growth, nodulation, yield and root-rot disease complex of chickpea under field condition. *African Journal of Biotechnology*. 2009;8(15).
14. Ali H, Khan MA, Randhawa SA. Interactive effect of seed inoculation and phosphorus application on growth and yield of chickpea (*Cicer arietinum* L.). *International Journal of Agriculture and Biology*. 2004;6(1):110-112.
15. Kumar SH, Dawson J, Kiran PS, Vyas VV. Effect of iron and zinc levels on growth and yield of Chickpea (*Cicer arietinum* L.) *Int. J. Curr. Microbiol. App. Sci*. 2020;9(11):2882-2886.
16. Bahure GK, Mirza IAB, Bankar AN, Puri AN, Sirsath MK. Effect of foliar application of zinc, iron and magnesium on growth, yield and quality of soyabean (*Glycine max* L.) Merrill. *Asian Journal of Multidisciplinary Studies*. 2016;4(3):19-22.
17. Mishra JP, Praharaj CS, Singh KK. Enhancing water use efficiency and production potential of chickpea and field pea through seed bed configurations and irrigation regimes in North Indian plains. *Journal of Food Legumes*. 2012b;25(4):310-313.
18. Lal B, Rana KS, Rana DS, Gautam, Priyanka, Shivay YS, Ansari MA, Meena BP, Kumar K. Influence of intercropping, moisture conservation practice and P and S levels on growth, nodulation and yield of chickpea (*Cicer arietinum* L.) under rainfed condition. *Legume Research*. 2014;37(3):300-305.
19. Kumar D, Arvadiya LK, Desai KL, Usadadiya VP, Patel AM. Growth and yield of chickpea (*Cicer arietinum* L.) as influenced by graded levels of fertilizers and bio fertilizers. *The Bioscan*. 2015;10(1):335-338.
20. Chavan S, Mansur CP, Shantveerayya. Soil moisture storage and productivity of chickpea as influenced by *in situ* moisture conservation practices in model watershed area. *Advances in Life Sciences*. 2016;5(3):1068-1073.
21. Gupta R, Chundawat GS, Aswani RC, Sarathe A. Performance evaluation of tractor operated furrow irrigated raised bed seed drill for Chickpea production. *Advances in. Bioresearch*. 2020;11(2):141-144.
22. Chandra R, Pareek N. Comparative performance of plant growth promoting rhizobacteria with rhizobia on symbiosis and yields in urdbean and chickpea. *Journal of Food Legumes*. 2015;28(1):86-9.
23. Gyandev, Kurdikeri BMB, Salimath PM. Effect of seed treatment on plant growth, seed yield and Quality of chickpea varieties. *International Journal of Agricultural Science and Research*. 2015;5(6):61-66.
24. Khaitov B, Kurbonov A, Abdiev A, Adilov M. Effect of chickpea in association with *Rhizobium* to crop productivity and soil fertility. *Eurasian Journal of Soil Science*. 2016;5(2):105-12.
25. Chauhan SVS, Singh RB. Effect of phosphorus and phosphate solubilizing bacteria on growth, yield and quality of chickpea (*Cicer arietinum* L.). *Annals of Plant and Soil Research*. 2017;19(3):303-306.
26. Singh AK, Chovatia PK, Kathiria RK, Savaliya NV. Effect of integrated nutrient management on growth, yield and economics of chickpea (*Cicer arietinum* L.). *International Journal of Chemical Science*. 2019;7(3):3048-3050.
27. Katiyar D, Kumar S, Singh N. Effect of *Rhizobium* and PSB inoculation on growth, yield attributes and yield of chickpea (*Cicer arietinum* L.). *International Journal of Chemical Studies*. 2020;8(4):3729-3734.
28. Benjelloun I, Thami Alami I, El Khadir M, Douira A, Udupa SM. Co-Inoculation of *Mesorhizobium ciceri* with Either *Bacillus* sp. Or *Enterobacter aerogenes* on Chickpea improves growth and productivity in phosphate-deficient soils in dry areas of a Mediterranean Region. *Plants*. 2021;10, 571.
29. Yadav A, Singh D, Kumar R, Sachan R, Kumar K, Singh A, Singh KK. Response of different level of phosphorus, zinc and *Rhizobium* inoculation on growth yield attributes and yield of Chickpea (*Cicer arietinum* L.). *International Journal of*

- Environment and Climate Change. 2022;12(11):1954-1964.
30. Gupta P, Samant K, Sahu A. Isolation of cellulose-degrading bacteria and determination of their cellulolytic potential. International Journal of Microbiology. 2012 Oct;2012.
 31. Yadav LR, Choudhary P, Santosh, Sharma OP, Choudhary M. Effect of phosphorus and zinc on yield and economics of mothbean under semi-arid conditions. Journal of Food Legumes. 2012;25(4):361-363.
 32. Singh AK, Bhati BP. Effect of foliar application of zinc on growth and seed yield of late-sown lentil (*Lens culinaris*). Indian Journal of Agricultural Sciences. 2013;83(6):622-626.
 33. Verma G, Yadav DD, Sharma VK, Kumar A, Singh RK, Upadhyay PK, Gupta G. Effect of fertility levels and biofertilizers on agrophysiological performance, productivity and quality of chickpea (*Cicer arietinum* L.). Indian Journal of Agricultural Sciences. 2019;89(9):1482–6.
 34. Verma G, Yadav DD, Kumar A, Singh R, Babu S, Avasthe RK, Gudade BA, Sharma VK. Impact of fertility levels and biofertilizers on root architecture, yield and nutrient uptake of Chickpea (*Cicer arietinum* L.). Int. J. Curr. Microbiol. App. Sci. 2020;9(2):2018-2024.
 35. Abisha P, Singh S. Effects of biofertilizer and phosphorus on growth, yield components and yield of Chickpea (*Cicer arietinum* L.). International Journal of Plant & Soil Science. 2022;34(20):326-331.
 36. Chaudhary, Seema, Singh H, Singh S, Singh V. Zinc requirement of green gram (*Vigna radiata*)-wheat (*Triticum aestivum*) crop sequence in alluvial soil. Indian Journal of Agronomy. 2014;59(1):48-52.
 37. Thenua OVS, Singh K, Vivek Raj, Singh J. Effect of sulphur and zinc application on growth and productivity of soybean [*Glycine max* (L.) Merrill] in Northern Plain Zone of India. Annals of Agricultural Research. 2014;35(2):183-187.
 38. Yadav P, Yadav DD, Pandey HP, Yadav A, Sachan R, Yadav S. Effect of fertility levels and biofertilizers on growth parameters, root architecture and quality of Chickpea (*Cicer arietinum* L.). International Journal of Plant & Soil Science. 2022;34(17):61-67.
 39. Kumar N, Singh MK, Praharaj CS, Singh U, Singh SS. Performance of chickpea under different planting method, seed rate and irrigation level in Indo-Gangetic Plains of India. Journal of Food Legumes. 2015;28(1):40-44.
 40. Singh Y, Singh B, Kumar A. Response of phosphorus levels and seed inoculation with PSB and *Rhizobium* on economic and response studies of Chickpea (*Cicer arietinum* L.) under rainfed condition. International Journal of Current Microbiology and Applied Sciences. 2017;6(11):801-805.
 41. Singh A, Singh G. Role of *Rhizobium* in Chickpea (*Cicer arietinum* L.) production. A review Agricultural Review. 2018;39(1):31-39.
 42. Singh A, Sachan AK, Pathak RK, Srivastava S. Study on the effects of PSB and *Rhizobium* with their combinations on nutrient concentration and uptake of chickpea (*Cicer arietinum* L.). Journal of Pharmacognosy and Phytochemistry. 2018;7(1):1591-1593.
 43. Singh AK, Meena MK, Bharati RC, Gade RM. Effect of sulphur and zinc management on yield, nutrient uptake, changes in soil fertility and economics in Rice (*Oryza sativa*)-lentil (*Lens culinaris*) cropping system. Indian Journal of Agricultural Sciences. 2013;83(3):344-348.
 44. Shivay YS, Prasad R, Madan Pal. Effect of variety and zinc application on yield, profitability, protein content and zinc and nitrogen uptake by chickpea (*Cicer arietinum*). Indian Journal of Agronomy. 2014;59(2):317-321.
 45. Parmar PM, Poonia TC, Raiyani VN. Agronomic biofortification of zinc in Chickpea varieties in calcareous soil. Legume Research; 2021. DOI: 10.18805/LR-4677

© 2022 Kumar et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/92373>