



Effect of *Cymbopogon citratus* Decoctions on Gasoline Vapour-induced Reproductive Toxicity in Female Rats

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Author's contribution

The sole author designed, analyzed and interpreted and prepared the manuscript.

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ABSTRACT

Aims: Recent research indicates that plant molecules, particularly those that are rich with antioxidant, anti-inflammatory and immune modulatory constituents, can modify and prevent the detrimental effects of gasoline compounds on reproductive endpoints. However, whether *C. citratus* decoction can alleviate gasoline vapour (GV)-induced derangement of female reproductive hormones has not yet been documented. In this study, the capacity of *C. citratus* decoction was evaluated for its ability to alleviate GV-induced reproductive toxicity in female rats.

Study Design: Seventy-two female Wistar albino rats weighing 185 ± 11.2 g were placed into six groups (n = 12 per group): the control (group 1, G1), GV alone (G2), GV plus *C. citratus* decoction (500 mg/kg; G3), (1000 mg/kg; G4), (1500 mg/kg; G5), and GV plus vitamin C (200 mg/kg; G6).

Place and Duration of Study: Department of Physiology, University of Uyo, Uyo, Akwa Ibom State, Nigeria. All groups were treated for 35 days.

Methodology: Serum levels of the female reproductive hormones progesterone (P₃) estradiol (E₂), luteinizing hormone (LH), and follicle stimulating hormone (FSH) as well as superoxide dismutase (SOD) and malondialdehyde (MDA; an oxidative stress marker) in the animals were assessed using standard procedures.

Results: The results showed that GV significantly ($p < 0.05$) decreased serum levels of P₃, E₂, LH,

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FSH, SOD and increased serum MDA levels compared to the levels in the control animals. However, co-administration of *C. citratus* at different doses to the animals in G3, G4, and G5 and vitamin C to the animals in G6 dose-dependently significantly ($p < 0.05$) increased the levels of the GV-reduced reproductive hormones and antioxidant enzyme and decreased the GV-increased oxidative stress marker levels to levels similar to those in the control group.
Conclusion: Thus, *C. citratus* decoction has an ameliorative effect on GV-induced reproductive dysfunction and oxidative stress.

Keywords: Attenuation; lemongrass; hydrocarbon; rats; reproductive toxicity.

1. INTRODUCTION

Gasoline is a mixture of several hydrocarbons, including aliphatics, aromatics, and several other branched saturated and unsaturated hydrocarbons, as well as many other additives. It is a volatile liquid and a ubiquitous hydrocarbon pollutant in the environment that is a particular problem for those living in close proximity to production, distribution, and retailing stations. Gasoline constituents particularly benzene, toluene, ethyl benzene, and xylene (BTEX) compounds are toxic to humans, and can affect many bodily functions [1,2].

Gasoline exposure is high among workers in oil exploration and retailing industries, and even in the general population during refuelling at self-service gasoline stations. In fact, it is estimated that 100 million people are exposed to gasoline constituents for a few minutes per week and approximately 100 minutes per year [3].

Interestingly, a toxic dose of gasoline can be inhaled within a very short period of time (seconds) [4]. It has been postulated that the air above an open barrel of gasoline in an unventilated out house on a "hot" day contains approximately 25,000 parts per million (ppm) of gasoline, the air around a tanker during bulk loading contains between 50–320 ppm, and the air around a petrol pump at service stations during vehicle refuelling contains 20–200 ppm [5,6].

Experimental and clinical data have established a close association between exposure to gasoline compounds and adverse reproductive endpoints [7,8,9], including changes in sexual behaviour, abnormal menstrual patterns and sex hormone profiles [10], decreased fertility, and interference with foetal development [11], leading to spontaneous abortion, birth defects, and intrauterine growth retardation [11]. Ugwoke et al. [8] reported structural disruptions in the uterus, fallopian tubes, and ovaries in female rats

and spermatocytic arrest, chromosomal abnormalities, and altered fertility in male rats acutely exposed to GV. Increased risks of cytogenetic alterations and mutagenic effects in both somatic cells and embryonic tissues have also been reported [12]. A recent study suggested that medicinal plant products could ameliorate gasoline compound (BTEX)-induced ovarian disorders. Indeed, gasoline compounds are regarded as environmental endocrine and reproductive toxicants/disruptors [13,14] that can modulate or interfere with the production, release, transport, metabolism, binding, actions, and elimination of natural hormones in the body [15,16].

Although the proposed molecular mechanisms through which GV induces reproductive toxicity may vary [17], including initiation of oxidative cell injury [18], suppression of rapidly growing cells (e.g. bone marrow cells by gasoline metabolites) [19,20], and impairment of hypothalamic-pituitary axis functions, accumulated data favour the induction of oxidative stress as the central pathophysiological mechanism [18].

Oxidative stress disturbs the antioxidant defence system, and through several intermediary mechanisms, it affects numerous bodily functions, including reproductive endpoints. Consequently, supplementation with antioxidants (synthetic or natural) may abate or reverse GV-induced reproductive toxicity. Several synthetic antioxidant minerals and vitamins, including vitamins A, C, and E, have been used to mitigate the deleterious effects of GV exposure in experimental models [21-24].

Accordingly, Uboh et al. [17] studied the effect of vitamin C on GV-induced reproductive toxicity and found that supplementation with vitamin C significantly reversed the reduced serum levels of sex hormones (progesterone and estradiol) to levels within the normal range and had an ameliorative effect on ovarian tissues in exposed

female rats. Their findings are consistent with several other studies in the literature [7-9].

Because of the inability of synthetic antioxidant monotherapies to address the pleiotropic effects of gasoline metabolites on the reproductive system and the drawbacks of multiple drug therapies, there is a fervent search for non-pharmacological means, specifically natural products, to reduce gasoline-induced reproductive toxicity and circumvent the limitations of multiple drug therapy. The numerous bioactive constituents present in natural products could mitigate the multi-dimensional effects of gasoline metabolites in gasoline-induced reproductive toxicity. This hypothesis is supported in a study conducted by Sirotkin et al. [2]. In that study, the authors found that administration of an extract from a medicinal plant (*Yucca*) restored ovarian cell function and attenuated the toxic effect of gasoline (BTEX) compounds. *Cymbopogon citratus* (*C. citratus*) is one such plant that has been explored for its antioxidant constituents and potentials [25].

C. citratus belongs to Poaceae family of plants and is widely distributed worldwide, especially in tropical and sub-tropical countries. It contains diverse bioactive constituents, including phytochemicals (saponins, tannins, flavonoids, phenols, anthraquinones, alkaloids, and deoxysugars) [26]; nutrients (proteins, moisture, ash, crude fibre, fat, and carbohydrates); electrolytes (potassium, calcium, copper, magnesium, manganese, selenium, phosphorus, iron, and zinc); and vitamins (folate, niacin, pyridoxine, and vitamins A, C, and E), and essential oil constituents (myrcene, neral, geraniol, linalool, limonene astral) [27]. Most of these constituents have recognized antioxidants anti-inflammatory and immune boosting effects.

However, whether *C. citratus* can ameliorate GV-induced reproductive toxicity has not yet been documented. Therefore, the present study aimed to accurately assess the effect of *C. citratus* decoction on gasoline-induced reproductive toxicity in female rats.

2. MATERIALS AND METHODS

2.1 Identification and Extraction

Fresh *C. citratus* leaves were obtained from an agricultural farm in Uyo, Akwa Ibom State, Nigeria a few days prior to utilization. Identification and authentication was performed

by a taxonomist in the Department of Botany at the University of Uyo. A voucher specimen (IDUHH4686/Uyo) was deposited in the herbarium for reference purposes. The leaves were rinsed, sundried, and pulverized into powder using an electric blender to provide 400 g of material. The powder was soaked with 4 L of hot water in a conical flask and allowed to stand for approximately 10 h. After filtering the solution through Whatman No. 2 filter paper, the filtrate was evaporated to dryness by heating in a water bath at 40°C. The final solid extract was weighed with an electric balance (ACS-2E14; Surgifriend Medicals, Ltd., England), with a total yield of 30%. The prepared extract was stored in glass bottles at 4°C and was dissolved in physiological saline at 100 mg/mL.

2.2 Phytochemical Screening of *C. citratus* Leaf Extract

The phytochemical analysis of the leaf extracts was carried out using standard procedures to determine the presence of tannins, phenolics, saponins, alkaloids, deoxysugars, and anthraquinones [28,29]. The concentrations of phytoconstituents were determined as described by Hajir et al. [30].

2.3 Experimental Animals

Seventy-two mature female Wistar albino rats weighing 185 ± 11.2 g were obtained from the animal house at the Faculty of Basic Medical Sciences, University of Uyo, Nigeria. The rats were divided into six groups with 12 rats per group, as follows:

- Group 1: Control group, unexposed
- Group 2: Test group exposed to GV only
- Group 3: Test group exposed to GV and concomitantly treated with *C. citratus* decoction (500 mg/kg; low dose) daily
- Group 4: Test group exposed to GV and concomitantly treated with (1000 mg/kg; medium dose) daily
- Group 5: Test group exposed to GV and concomitantly treated with (1500 mg/kg; high dose) daily
- Group 6: Test group exposed to GV and concomitantly treated with vitamin C (200 mg/kg)

2.4 Determination of LD50

The median lethal dose (LD50) was determined using standard methods [31]. From the LD50

(5000 mg/kg), the low, medium, and high doses were calculated using a standard formula as 500 mg/kg, 1000 mg/kg, and 1500 mg/kg, respectively. The rats were acclimatized for one week before the starting the experiment. Each animal was housed in a standard wooden cage with wood shavings as bedding, which was regularly replaced. The experiment was performed under standard laboratory conditions (at room temperature [$28 \pm 8^\circ\text{C}$], 45% humidity, with a 12-h light/dark cycle). All animals were fed normal rat pellets and allowed free access to food and water throughout the experimental period.

All research protocols were performed at the University of Uyo according to Nigerian and international laws governing the acceptable use of laboratory animals.

2.5 Exposure to GV

The animals in the test group were exposed to unleaded gasoline purchased from a Nigerian National Petroleum Cooperation (NNPC) refuelling station on Itam-Ikot Ekpene Road in Uyo, Nigeria. The rats in the exposed groups were housed in their cages and exposed to GV in an exposure chamber (80 x 60 x 100 cm). Rats in the control group were kept in a GV-free section of the experimental house. Two calibrated 500-mL beakers each containing 100 mL of petrol were put in the modified chamber where the exposed groups were placed, and then the rats were allowed to inhale the GV in the chamber for 6 h (9 am–3 pm) daily for 35 consecutive days. At the end of the exposure period, the gasoline was removed; the initial and final volumes were recorded before and after exposure, respectively. The daily difference in volume was used to estimate the relative vapour exposure. The average exposure was approximately 80 mL/day. Rats in groups 3, 4, and 5 were treated with decoctions prepared from 500 mg/kg, 1000 mg/kg, and 1500 mg/kg of *C. citratus* decoction, respectively, while rats in group 6 were treated with 200 mg/kg (i.e. the normal prophylactic dose) of vitamin C [17]. The *C. citratus* decoctions and water-solubilized vitamin C were administered by oral gavage (using an intragastric syringe) for the final 14 days of the 35-day GV exposure.

2.6 Biochemical Assay

After the exposure period, the rats were sacrificed by anaesthetizing with chloroform.

Blood was collected by cardiac puncture for biochemical analyses, including measurement of serum female reproductive hormone profile (progesterone [P3], estradiol [E2], follicle stimulating hormone [FSH], and luteinizing hormone). Serum malondialdehyde (MDA) and superoxide dismutase (SOD) levels were determined using the double heating method of Draper and Hadley as described elsewhere [32].

2.7 Statistical Analysis

Statistical analysis was performed using the Statistical Package for Social Sciences (SPSS), version 20.0. Data obtained were analysed using descriptive statistics and reported as the mean \pm standard deviation (SD). Analysis of variance (ANOVA) was also used. Duncan's multiple range test was used to compare results between groups, determine the direction of significance, and to analyse the effect of the *C. citratus* decoction on the reproductive toxicity of GV. Differences with p values less than 0.05 were considered statistically significant.

3. RESULTS AND DISCUSSION

The result of this study showed that exposure to gasoline compounds is a significant risk for reproductive function impairment, and that, the use of plants and plant products, particularly those with potent antioxidant, anti-inflammatory and immune modulatory compounds (such as *C. citratus*) can alleviate GV-induced adverse reproductive endpoints. Evidence for this contention is twofold: first, the significant decreases in the serum levels of reproductive hormones (E_2 , P_3 , LH, and FSH) and antioxidant enzyme (SOD) and a significant increase serum MDA in animals exposed to GV alone compared to the corresponding levels in control animals; second, the reversal of these changes following the co-administration of *C. citratus* leaf decoction at 500 mg/kg, 1000 mg/kg, and 1500 mg/kg and vitamin C to animals in groups 3, 4, 5 and 6, respectively.

These results confirm previous speculation that gasoline compounds down regulate ovarian steroid hormones levels via several unexplained mechanisms, including inhibition of increased hypothalamic GnRH/pituitary gonadotropin production [17] as evidence by the significant decrease in E_2 , P_3 , LH, and FSH levels.

These observations corroborate previous studies showing that inhalation of toluene by rats was

associated with a reduction in the hypothalamic levels of gonadotropins and alteration of FSH and LH. Likewise, xylene exposure led to decrease in plasma P₃ and E₂. Oxidative cell injury is the postulated pathophysiologic mechanism underlying gasoline compound-induced reproductive toxicity [18]. For example, one gasoline compound, toluene, has been shown to decrease glutathione peroxidase activity and increase lipid peroxidation in ovarian tissues, resulting in ovarian cell damage [18]. Therefore, administration of antioxidant substances (synthetic or natural) could inhibit or even reverse the toxic effects of gasoline-induced oxidative stress and subsequent reproductive dysfunction. This hypothesis is supported in studies by Uboh et al. [17] and Sirotkin et al. [2], which showed that vitamin C had a ameliorative effect on GV-induced impaired reproductive function in rats, and that Yucca extract modified and abated the effect of benzene on ovarian hormone release, respectively.

Our results showed increased oxidative stress in animals exposed to GV alone as evidenced by a significant increase in MDA which is a common marker of lipid peroxidation [33] and decrease in antioxidant enzyme (SOD) levels, evidence of oxidative stress. Elevated MDA indicates peroxidative damage to cell membranes and other lipid-derived macromolecules [4]. Similar to previous studies [4], an increase in the lipid peroxidation product (MDA) was correlated with decrease in superoxide dismutase (SOD). This creates an imbalance between pro-oxidants and antioxidants, which has been shown to be associated with adverse female reproductive endpoints [34], including a reduction in the reproductive hormone profile [35]; increased risk of abortion; low birth weight [35], and intrauterine growth retardation (IUGR) [11]. Others include structural disruption in the uterus, fallopian tubes, and ovaries [8]; disordered menstrual pattern [10]; ovarian hypo and hyperplasia [36,37]; and reduced hypothalamic levels of gonadotropin releasing hormone (GnRH) and plasma gonadotropin levels [2,38].

In support of this observation, Su et al. [39] have also reported that some medicinal plants products, either alone or in combination, have the potential to restore reproductive organ degeneration by improving the histo-morphology of the ovaries, uterus, and vagina and modulating E₂ imbalance and neurotransmitter disorders.

Likewise other studies have shown that medicinal plant products can abate gasoline (BETX) compounds-induced ovarian dysfunction, including changes in the release of P₃, insulin growth factor-1 (IGF-1), and prostaglandin F (PGF), and prevent the effects of benzene on ovarian hormone (P₃ and E₂) release.

C. citratus and vitamin C have recently received attention as natural and synthetic antioxidants, anti-inflammatory and immune modulators respectively, with great potential to alleviate drug/chemical-induced oxidative stress, inflammation and immune perturbation and associated disorders [17,25,40-42]. In a study by Rahim et al. [41] to evaluate the effect of *C. citratus* aqueous extract and vitamin E on hydrogen peroxide-induced reproductive function impairment in male rats, *C. citratus* significantly increased the body, testicular and epididymal weight, and testosterone and GSH levels. Improvement in values of semen characteristics and reduction in serum and testicular MDA levels were also observed. Supplementation with the *C. citratus* aqueous extract also caused improvement in testicular histopathological alteration. The antioxidant activity of *C. citratus* extract is due to its constituent phytochemicals (saponins, tannins, polyphenols, and flavonoids), vitamins (vitamins A, C, and E), minerals and trace elements (Zn, Cu, Mg, Se, and Fe), and essential oil components (geraniol, myrcene, linalool, limonene, β-ionone, and citral) [43] (Table 1).

Some of these constituents provide direct antioxidant effects, while others, such as Se, Cu, Mn, and Zn, do not have antioxidant effects themselves, but are required for the activity of some antioxidant enzymes [32]. Vitamin C acts as a first-line antioxidant in plasma [44]. It inhibits LDL oxidation and maintains normal endothelial function. The rich antioxidant nutrient composition of *C. citratus* has been exploited for the management of several drug/chemical-induced oxidative stress-mediated adverse health effects including cisplatin, carbon tetrachloride, and hydrogen peroxide-induced liver damage [25,39,40] respectively, and isoprenoid-induced cardiotoxicity and lipid peroxidation [45] in rats. Here we showed that *C. citratus* leaf extract has an ameliorative effect on GV-induced reproductive system dysfunction, partly due to its rich antioxidative anti-inflammatory and immune modulatory constituents and their activities (Table 2).

Table 1. Phytochemical constituents of *C. citratus* leaf extract

Composition	Tannins	Saponins	Flavonoids	Alkaloids	Phenols	Anthraquinones
Concentration	++	++	++	+	++	+

Key: + Present, ++ Moderate

Table 2. Effect of *C. citratus* decoctions on GV-induced derangement in serum female sex hormone profile

Groups	FSH (m μ /mL)	LH (m μ /mL)	Estradiol (ng/mL)	Progesterone (ng/mL)	SOD (mmol/L)	MDA (mmol/L)
G1 (control)	5.14 \pm 2.02 ^b	13.12 \pm 2.47 ^b	56.22 \pm 6.72 ^c	36.08 \pm 8.22 ^c	10.530 \pm 1.240 ^b	5.81 \pm 0.55 ^a
G2	2.32 \pm 1.04 ^a	8.06 \pm 3.14 ^a	25.24 \pm 5.22 ^a	16.02 \pm 9.11 ^a	7.064 \pm 1.520 ^a	11.10 \pm 0.41 ^c
G3	3.08 \pm 0.99 ^b	10.14 \pm 2.02 ^b	28.38 \pm 18.20 ^a	20.82 \pm 6.54 ^a	7.840 \pm 1.380 ^a	9.03 \pm 0.45 ^c
G4	3.86 \pm 0.52 ^b	10.84 \pm 1.82 ^b	32.42 \pm 6.47 ^b	26.42 \pm 5.85 ^b	8.24 \pm 2.24 ^a	7.42 \pm 0.39 ^b
G5	4.06 \pm 1.22 ^b	11.24 \pm 1.22 ^b	38.06 \pm 3.04 ^b	28.26 \pm 4.61 ^b	9.76 \pm 2.81 ^b	7.07 \pm 0.39 ^b
G6	4.16 \pm 0.78 ^b	12.06 \pm 3.72 ^b	40.42 \pm 4.82 ^b	30.22 \pm 5.88 ^c	10.23 \pm 2.88 ^b	4.18 \pm 3.27 ^a

Similar letters/alphabet mean not significantly different (p > 0.05),

Different letters mean significantly different (p < 0.05), Values are reported as mean \pm SD.

FSH: follicle stimulating hormone; LH: luteinizing hormone; SOD: superoxide dismutase; MDA: malondialdehyde

4. CONCLUSION

Results of the present study indicate that *C. citratus* decoction has an ameliorative effect on GV-induced reproductive dysfunction and oxidative stress.

CONSENT

It is not applicable.

ETHICAL APPROVAL

The author hereby declares that "Principles of laboratory animal care" (NIH publication No. 85-23, revised 1985) were followed, as well as specific national laws where applicable. All experiments have been examined and approved by the appropriate ethics committee.

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COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

1. Azari MR, Konjin ZN, Salehpour FZ, Seyedi MD. Occupational exposure of petroleum depot workers to BTEX compounds. *Int J Occup Environ Med.* 2012;3:39-44.
2. Sirotkin V, Kadasi A, Balazi A, Bakova Z, Harrath AH, Makarevich AV, et al. Influence of petrochemical industry environmental contaminants on animal ovarian cells. *J Microbiol Biotech Food Sci.* 2012;2:517-525.
3. Caprino, Togna GI. Potential health effects of gasoline and its constituents: A review of current literature (1990-1997) on toxicological data. *Environ Health Perspect.* 1998;106:115-125.
4. Odewabi O, Ogundahunsi OA, Oyalowo M. Effect of exposure to petroleum fumes on plasma antioxidant defense system in petrol attendants. *Br J Pharmacol Toxicol.* 2014;5:83-87.
5. Micyus J, MuCurry JD, Seeley JV. Analysis of aromatic compounds in gasoline with flow switching comprehensive two-dimensional gas chromatography. *J Chromatogr.* 2005;1086: 115-121.
6. Lewne M, Nise G, Lind ML, Gustavsson P. Exposure to particles and nitrogen dioxide among taxi, bus drivers. *Int Arch Occup Environ Health.* 2006;79:220-226.

7. Ugwoke C, Nwobodo E, Unekwe P, Odike M, Chukwuma S. Fuel vapour disrupts the histo-architecture of reproductive organs in albino rats. *J Sci Ind Studies*. 2004;2:72-74.
8. Ugwoke C, Nwobodo ED, Unekwe P, Odike M, Chukwuma ST, Amilo G. The reproductive dysfunction effects of gasoline inhalation in albino rats. *Nig J Physiol Sc*. 2005;20:54-57.
9. Uboh E, Akpanabiatu MI, Ekaidem IS, Ebong PE, Umoh IB. Effect of inhalation exposure to gasoline fumes on sex hormones profile in Wistar albino rats. *Acta Endocrinol (Buc)*. 2007;4:23-30.
10. Ekpenyong C, Davies K, Daniel N. Effects of gasoline inhalation on menstrual characteristics and the hormonal profile of female petrol pump workers. *J Environ Protect*. 2013;4:65-73.
11. Slama O, Thiebaugeorges V, Goua L, Aussel P, Sacco A, Bohet, et al. Maternal personal exposure to air born benzene and intrauterine growth. *Environ Health Perspect*. 2009;177:1313-1321.
12. Hatch C, Warburton D, Santella RM. Polycyclic aromatic hydrocarbon-DNA adducts in spontaneously aborted fetal tissues. *Carcinogenesis*. 1990;11:1673-1675.
13. Humfrey ON, Smith LL. Endocrine disrupting chemicals. The evidence for human health effects in endocrine and hormonal toxicity. Ed. Harvey PW, Rush KC, Cockborn A. (John Wiley and Son, NY, USA). 1999;421.
14. Mattison DR, Plowehalk DR, Meadow MJ, Al-juburi AZ, Gardy J, Malek A. Reproductive toxicity: Male and female reproductive systems and targets for chemical injury. *Med Clin North Am*. 1990;74:391-411.
15. Kavlock J, Daston GP, DeRosa C, Fenner-Crisp P, Gray LE, Kaattari S, et al. Research needs for risk assessment of health and environmental disruptors: A review of U.S. EPA-Sponsored workshop. *Eviron Health Perspect*. 1998;104:715-740.
16. Nicolopoulou-Stamati P, Pitsos MA. The impact of endocrine disruptors on the female reproductive system. *Human Reproduct Update*. 2000;7:323-330.
17. Uboh F, Ebong P, Udosen E. Effect of vitamin C on gasoline vapor induced reproductive toxicity in female rats. *Turk J*. 2010;14:26-30.
18. Burmistrov SO, Arutyunyan AV, Stepanov MG, Oparina TI, Prokopenko VM. Effect of chronic inhalation of toluene and dioxane on activity of free radical processes in rat ovaries and brain. *Bull Exp Biol Med*. 2001;132(3):832-836.
19. Rao NR, Snyder R. Oxidative modifications produce in HL-60 cells on exposure to benzene metabolites. *J Appl Toxicol*. 1995;15:403-409.
20. Laskin D, Rao NR, Punjabi CJ, Laskin DL, Synder R. Distinct actions of benzene and its metabolites on nitric oxide production by bone marrow leukocytes. *J Leukoc Biol*. 1995;57:422-426.
21. Ayo O, Minka NS, Mamman MM. Excitability scores of goats administered ascorbic acid and transported during hot-dry conditions. *J Vet Sci*. 2006;7:127-131.
22. Ambali S, Akanbi D, Igbokwe N, Shittu M, Kawu M, Ayo J. Evaluation of sub-chronic chlorpyrifos poisoning on haematological and serum biochemical changes in mice and protective effect of vitamin C. *J Toxicol Sc*. 2007;32:111-120.
23. Bashandy A, Alhazza IM. The hepatoprotective effect of α -carotene against cadmium toxicity in rats. *J Pharmacol Toxicol*. 2008;3:457-463.
24. Uboh E, Akpanabiatu MI, Alozie Y. Comparative effect of gasoline vapor on renal functions in male and female Wistar rats. *J Pharmacol Toxicol*. 2008;3:478-484.
25. Koh H, Mokhtar RA, Igbal M. Antioxidant potential of *Cymbopogon citratus* extract: Alleviation of carbon tetrachloride-induced hepatic oxidative stress and toxicity. *Hum Exp Toxicol*. 2012;31:81-91.
26. Ekpenyong CE, Daniel NE, Antai AB. Bioactive natural constituents from lemongrass tea and erythropoiesis boosting effects: Potential use in prevention and treatment of anemia. *J Med Food*. 2015;18:118-127.
27. Shah G, Shri R, Panchai V, Sharma N, Singh B, Mann AS. Scientific basis for the therapeutic use of *Cymbopogon citratus* stapf (Lemongrass). *J Adv Pharm Technol Res*. 2011;2:3-8.
28. Trease E, Evans C. A textbook of pharmacognosy. 13th Ed., Baluire, Tindali, London. 1989;100-101.
29. Sofowora A. Medicinal plant and traditional medicine in Africa, Second ed., Spectrum Books, Ibadan, Nigeria. 1996;112.

30. Hajir BA, Alaa IM, Khansa AA, Naga IA, Wdeea A, Ahmed AE, Ahmed JA, Ismail GM, Anas ME, Nagat AE, Samir FA, Monier NH. Evolution of antimicrobial, antioxidant potentials and phytochemical studies of three solvent extracts of five species from acacia used in Sudanese ethnomedicine. *Advances in Microbiology*. 2016;6:691-698.
31. Lorke D. A new approach to practical acute toxicity testing. *Arch Toxicol*. 1983;53: 275-287.
32. Ekpenyong C, Bassey G. Investigation of the effect of *Cymbopogon citratus* decoctions on gasoline vapour-induced lipid peroxidation and hepatotoxicity in rats. *J Int Res Med Pharm Sci*. 2016;10(4): 171-181.
33. Agarwal A, Chandra G, Singh AK. Heat shock protein 70, oxidative stress and antioxidant status in periparturient crossbred cows supplemented with α -tactopherol acetate. *Trop Anim Health Prod*; 2012.
DOI: 10.1007/s11250-012-0196-z
34. Al-Gubbory H, Fowler PA, Garrel C. The roles of cellular reactive oxygen species, oxidative stress, and antioxidants in pregnancy outcomes. *Int J Biochem Cell Biol*. 2010;42:1634-1650.
35. Hakkola M, Honkasalo ML, Pulkkinen P. Neuropsychological symptoms among tanker drivers exposed to gasoline. *Occup Med*. 1996;46:125-130.
36. Chen D, Cho S, Chen C, Wang X, Damokosh AI, Ryan L, et al. Exposure to benzene, occupational stress, and reduced birth weight. *Occup Environ Med*. 2000;57:661-667.
37. Maronpot R. Ovarian toxicity and carcinogenicity in eight recent National Toxicity Program Studies. *Environ Health Perspect*. 1987;73:125-130.
38. Hannigan H, Bowen ES. Reproductive toxicology and teratology and abused toluene. *Sys Biol Reprod Med*. 2010;56:184-200.
39. Su JY, Xie QF, Liu WJ, Lai P, Liu DD, Tang LH, Dong TT, Su ZR, Tsim KW, Lai XP, Li KY. Perimenopause Amelioration of a TCM recipe composed of radix astragali, radix angelisae, sinensis, and folium epimedi: An *in vivo* study on natural aging rat model. *Evidence-based Complement Altern Med*; 2013.
DOI: org/10.1155/2013/747240
40. Arhoghro M, Kpomah DE, Uwakwe AA. Curative Potential of Aqueous Extract of Lemon Grass (*Cymbopogon citratus*) on Cisplatin Induced Hepatotoxicity in Albino Wistar Rats. *J Physiol Pharmacol Adv*. 2012;282-294.
41. Rahim SM, Taha EM, Al-janabi MS, Al-douri BI, Simon KD, Mazian AG. Hepatoprotective effect of *Cymbopogon citratus* aqueous extract against hydrogen peroxide-induced liver injury in male rats. *Afri J Tradit Complement Altern Med*. 2014;11:447-451.
42. Uboh E, Akpanabiatu M, Edet E, Ebong P. Vitamin A, C and E ameliorate gasoline vapour-induced toxicity effect on sex hormoneal levels in female Wistar Rats. *Gen Endocrinol*. 2011;7:189-198.
43. Saito ML, Sramin S. Plantas aromáticas eseu uso na agricultura. *Jaguariúna: Embrapa*, 2000;45.
44. Frei B, England L, Ames BN. Ascorbate is an outstanding antioxidant in human blood plasma. *Proc Nat Acad Sci USA*. 1989;86:6377-6381.
45. Gayathri K, Jayachandran KS, Vasanthi HR, Rajamanickam GV. Cardio-protective effects of lemongrass as evidence by biochemical and histo-pathological changes in experimentally induced cardio-toxicity. *Human Exp Toxicol*. 2011;30(8):1073-1082.

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