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Experimental Investigation of the Functional and Proximate Properties of Pigeon Pea (*Cajanus cajan*) Using Fermentation Process

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Authors' contributions

This work was carried out in collaboration among all authors. Author BFO designed the study, supervised the project and wrote the first draft of the manuscript. Authors JAI and OJO contributed to the literature search, method of the research and corrected the final manuscript authors OEA and ASO carried out the processing, laboratory analysis and managed the analyses of the study. All authors read and approved the final manuscript.

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Original Research Article

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ABSTRACT

Pigeon pea (*Cajanus cajan*) is a leguminous crop rich in protein and widely cultivated in Nigeria but underutilized due to high anti-nutritional properties. Several processing methods such as fermentation using non-pathogenic micro fungi have been used to address these problems. This study evaluated the effect of fermentation on the functional and proximate properties of pigeon pea using *Rhizopus* species. The following ranges of values were obtained for bulk density, water absorption, oil absorption and swelling capacities (0.74-0.98g/ml; 71.67-189.00%; 120.00-205.00%; 6.81-10.93g/ml), respectively. Values of proximate composition were in ranges of 6.67-7.87, 16.09-22.09, 2.83-4.00, 0.84-1.23,1.33- 2.00 and 65.47- 71.35% for moisture, protein, ash, fat, fibre and carbohydrate contents, respectively. Improvement in functional and proximate properties of fermented pigeon pea flour were observed. Hence, it can also serve as functional and supplement ingredients in different food formulations.

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Keywords: Pigeon pea; fermentation; water absorption; bulk density; Rhizopus.

1. INTRODUCTION

Pigeon pea is recognised as perennial legume having good nutritional values that can be used in the management of metabolic problems in and animals According human [1]. to Venkidasamy et al. [2], pigeon peas is a typical legume rich in protein and are served as staple food for large population in India. This accounts for India being the largest supplier of the pigeon pea in the world market [3]. The rate of pigeon pea intake thus has contributed to consumption of reduced fat. and consequently helping in building the body weight [4].

Despite the potentials of pigeon pea, there seems to be problems associated with its processing making it impossible for consumers to adequately subscribe to its consumption. For instance, Feng et al. [5] discovered that certain methods for its processing pose some quality defects such as quality loss in the food, taste, flavour and colour to make it unhealthy for consumption. However, optimization of the fermentation parameters gives a better quality of the pigeon pea. More so, Lee et al. [6] reported that fermented pigeon pea improves the regulation of blood pressure which helps in reducing cardiovascular health challenges. Based on this, Olagunju et al. [7] hydrolysed pigeon pea protein using different enzymes and observed that membrane ultrafiltration exists which has the potential of improving the functional recipes in industry in relation to plant based products.

Many researchers focused on utilizing the pigeon pea towards further enhancement of the nutritional value of some foods. For instance, study by Solomon et al [8] revealed that toasted pigeon pea can be used to improve nutritional quality of cat fish, thereby reducing the cost of catfish production. In view of this, Martinez-Villaluenge et al. [9] further applied fermented Cajanus cajan powder to improve the quality of protein in pasta. Further studies on pigeon pea showed that different modification methods have effects on both functional and proximate properties to consequently influence the variation in their suitability for varieties of products that can be developed [10-15]. Thus, the aim of this study is to investigate effect of fermentation on the functional and proximate properties of pigeon pea (Cajanus cajan).

2. MATERIALS AND METHODS

2.1 Materials

Pigeon pea seeds were obtained from a local market in Ogbomoso while the *Rhizopus species* were obtained from Microbiology Laboratory of Food Science and Engineering Department LAUTECH, Ogbomoso.

2.2 Methods

2.2.1 Preparation of subculture for pigeon pea fermentation

Rhizopus oligosporus, Rhizopus oryzae and Rhizopus nigricans were sub cultured according to the procedure of Olanipekun et al. [16]. Five hundred millilitre (500ml) of Potato Dextrose Agar (PDA) was obtained through dissolution of 18 g of PDA to 500 ml of distilled water, homogenised and subsequently sterilized. After cooling, 15 ml of PDA was dispensed into McCartney bottles, and allowed to set in slant form. The pure cultures of Rhizopus oligosporus, Rhizopus oryzae and Rhizopus nigricans were sub cultured singly into McCartney bottles and incubated at 30°C for 4 days.

2.2.2 Procedure of pigeon pea fermentation

The pigeon pea seeds were cleaned, steeped $(28 \pm 2 \degree C \text{ for } 24 \text{ h})$, dehulled, boiled $(100 \degree C \text{ for } 15 \text{ min})$, drained and cooled. Subsequently, the seeds were inoculated using different Rhizopus species (*Rhizopus oryzae, Rhizopus oligosporus* and *Rhizopus nigricans*), sealed in polyethylene bag, incubated (32 \degree C for 72 h), pulverized, dried (55 °C for 24 h), cooled, milled and sieved to produce fermented pigeon pea flour. The unfermented pigeon pea (0 h fermentation) serves as the control and at regular intervals of 12 h; sample was taken out for appropriate analysis.

2.3 Analysis of the Samples

Water absorption capacity was carried out using the procedure of Owuamanam *et al.* [17] and bulk density was determined by using the method of Nwanekezi *et al.* [18]. Swelling capacity of the samples was determined by Tester and Morrison [19]. The proximate composition was determined by the method described by AOAC [20] while carbohydrate was calculated by difference. Oil absorption capacity was determined using the method of Adelakun *et al.* [21]. All the analyses were performed in triplicates.

3. RESULTS AND DISCUSSION

3.1 Functional Properties of Fermented Pigeon Pea

The results of functional properties of fermented pigeon pea presented in Table 1 showed decreasing trend in water absorption of pigeon pea fermented using different species of Rhizopus nigricans, Rhizopus orvze and Rhizopus oligosporus singly within 24-72 h, respectively. The trend in water absorption was observed to follow decreasing pattern of 189.00-74.67, 179.00 -71.67 and 174.00-73.33%, respectively. Consequently, the findings are in agreement with the studies by Olanipekun et al. [22], and Oyelade et al. [23] as water absorption was found to decrease for cocoyam and soya beans due to fermentation.

Swelling capacity exhibited slight decrease in their values which ranged as 8.27- 6.81, 10.93 -7.11 and 10.74- 7.23 g/ml, respectively using *Rhizopus nigricans, Rhizopus oryzae* and *Rhizopus oligosporus.* Fermenting pigeon pea with *Rhizopus oryzae* gave the highest value of 10.93 g/ml whereas the least value of 6.81 g/ml was obtained using *R. nigricans* at 72 h of fermentation period. There were significant differences between the samples (p<0.05). It was observed that swelling capacity decreased with increase in fermentation period. The result obtained is in agreement with the report of Lawal *et al.* [24] on effects of selected Hofmeister anions on functional properties of protein isolate prepared from lablab seeds. Adebowale and Maliki [25] found similar trend on the chemical composition and functional properties of fermented pigeon pea (*Cajanus cajan*) flour. Variation in swelling power observed could be due to effect of particle size and nature of unmodified starch in the pigeon pea as probably affected by fermentation which invariably may affect stickiness.

The oil absorption capacity of fermented flour decreased as the fermentation period increases except when using *Rhizopus nigricans* that showed contrary trend. The values ranged from 172.00-183.00, 199.00 -194.00 and 205.00-120.00%, respectively using *Rhizopus nigricans*, *R.oligosporus* and *Rhizopus oryzae*. The control sample had the value of 185.00%. Fermenting pigeon pea with *Rhizopus oligosporus* for 48 h gave the highest value while the least value obtained when using *Rhizopus oryzae* at 72 h. The decreasing trend agreed with the report of Adebowale and Malaiki [25] on the pigeon pea flour.

Bulk density of fermented pigeon pea increased as the fermentation period increases. This is indicated in the values 0.74 – 0.98, 0.77-0.98 and 0.92-0.98g/ml obtained when using *Rhizopus nigricans*, *Rhizopus oligosporus* and *Rhizopus oryzae*, respectively. The highest value was obtained in samples C, F and I while the least value obtained in sample A. Fermenting

Table 1. Functional	properties of	f fermented	pigeon p	bea
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Sample	Water Absorption Capacity (%)	Swelling Capacity (g/ml)	Oil Absorption Capacity (%)	Bulk Density (g/ml)
Α	189.00±4.16 ^a	8.27±0.500°	172.00±7.211 ^d	0.74±0.000 ^d
В	194.00±5.774 ^a	8.19±0.388°	162.00±1528 ^e	0.94±0.025 ^b
С	74.67±1.528 ^e	6.81±0.799 ^e	183.00±8.622°	0.98±0.027ª
D	179.00±5.686 ^a	10.93±0.397ª	199.00±8.083 ^a	0.77±0.023 ^d
E	170.00±1.824 ^c	8.94±1.101 ^b	208.00±1.000 ^a	0.93±0.025 ^b
F	71.67±2.517 ^e	7.11±0.549 ^e	194.00±9.292 ^a	0.98±0.027ª
G	174.00±2.539 ^b	10.27±0.450 ^a	205.00±10.066 ^a	0.92±0.122°
Н	154.00±2.000 ^d	10.74±0.214ª	184.00±15.044°	0.95±0.000 ^a
I	73.33±3.786 ^e	7.23±0.387 ^d	120.00±7.506 ^f	0.98±0.027ª
J	66.33±0.577 ^f	11.93±0.809 ^a	185.00±4.993 ^{bcd}	0.90±0.023 ^b

Sample A, B and C: Pigeon pea fermented with Rhizopus nigricans at 24, 48 and 72 h, respectively Sample D, E and F: Pigeon pea fermented with Rhizopus oligosporus at 24, 48 and 72 h, respectively Sample G, H and I: Pigeon pea fermented with Rhizopus oryzae at 24, 48 and 72 h, respectively Sample J: Control sample pigeon pea using Rhizopus nigricans at 24 h gave the least value compared to the highest values obtained at 72 h using Rhizopus nigricans, Rhizopus oligosporus and Rhizopus oryzae, respectively. The bulk density values obtained in this study were higher than those of Edema et al. [26] obtained for soybean flour (0.38g/ml). However, the result of the findings is in agreement with the reports of Onimawo et al. [27] which had the values that ranged from 0.06 and 0.60-0.75g/ml for cowpea and bambara groundnut. respectively.

3.2 Proximate Composition of Fermented Pigeon Pea

The result of proximate composition of fermented pigeon pea flour using Rhizopus nigricans, Rhizopus oligosporus and Rhizopus oryzae is presented in Table 2. The level of protein content increased from 17.10 - 22.09, 18.27 - 20.13 and -18.49% using 16.71 Rhizopus nigricans, Rhizopus oligosporus and Rhizopus oryzae, respectively. Fermenting pigeon pea with Rhizopus nigricans gave the highest value of 22.09% while the least value of 16.09% was obtained in an unfermented sample. The increase in protein content is in agreement with the results obtained during fermentation of beniseed flour and pearl millet according to Oshodi et al. [28]; Fasasi [29].

The ash content of the fermented pigeon pea showed decreasing trend from 3.17 - 3.00, 3.67 -2.67 and 3.50-2.83% using Rhizopus nigricans, Rhizopus oligosporus and Rhizopus oryzae, respectively. Fermenting pigeon pea with Rhizopus. oligosporus gave the highest value of 4.00% whereas sample F fermented using Rhizopus oligosporus at 72 h gave the least value of 2.67%. Increment in the ash content of fermented pigeon pea was in accordance with the reports of SefaDedeh et al. [30]; Adebowale and Maliki [25] on the fermentation of cowpea and fortification for the production of traditional foods, respectively. This similar finding was reported by Emmanuel et al. [31] for locust bean (Parkia biglobosa).

The moisture content of pigeon pea fermented using *R. nigricans* increased from 6.90-7.20 % whereas others slightly decreased from 7.20-7.13% and 7.30 -6.69 % using *Rhizopus oligosporus* and *Rhizopus oryzae, respectively.* Fermenting pigeon pea using *Rhizopus oligosporus* and *Rhizopus oryzae* which indicated decreasing trend is good for enhanced keeping quality of product. Similar trend has been observed on complementary food formulated from fermented maize, soybean and carrot water as reported by Lucretia *et al.* [32].

Fat content of the fermented pigeon pea flour showed a decreasing trend ranging from 1.10 -0.80, 1.01-0.91 and 0.92- 0.84% using Rhizopus niaricans. Rhizopus oligosporus, R.oryzae, respectively. The control sample had the highest value of 1.23% while sample C having the lowest value of 0.80%. Slight differences were observed in all samples at p<0.05. The result obtained is in agreement with the findings of Adebowale et al.,[33]; Anthony and Babatunde [34]; Emmanuel et al. [31] on fermented legumes and cereals. content observed low-fat has The potential to improve shelf-life through rancidity reduction.

The carbohydrate content of fermented pigeon pea flour showed a slight decreasing trend from 67.78 -65.47, 67.86- 67.40 and 69.81-68.56% using Rhizopus nigricans, Rhizopus oligosporus and Rhizopus oryzae, respectively. Sample J had the highest value of 71.35 % and Sample C had the lowest values of 65.47%. Slight differences were observed in all the samples at p<0.05. This result obtained is in accordance with the findings of Emmanuel et al. [31]; Ahmed et al. [35] during fermentation of locust bean =and guar seeds. The decreasing trend was attributed to increase activity of amylolytic enzymes which hydrolyses starch and other complex carbohydrate to simpler starch (Adebowale and Maliki, [25]; Ojokoh et al., [36].

Fiber content of the fermented pigeon pea flour showed decreasing values of 1.65 - 1.50, 2.00-1.33 and 1.75-1.42% using Rhizopus nigricans, Rhizopus oligosporus and Rhizopus oryzae respectively. Sample D had the highest value of 2.00% and sample F had the lowest value of 1.33%. Generally, the decreasing trend in fiber content could be linked to the observation of Omafuvbe et al. [37] indicating that at the end of boiling process, the boiled water obtained from the beans was observed to be more viscous than it was at the beginning of the process. The result is also similar to the finding of Onweluzo [38] that reported decrease during the fermentation of millet and pigeon pea seed for flour production.

Sample	Moisture Content (%)	Protein %	Ash (%)	Fat (%)	Fibre (%)	Carbohydrate (%)
А	6.90±0.529ª	17.10±1.186 ^a	3.17±0.289 ^b	1.10±1.123 ^{ab}	1.65±0.132 ^{abc}	67.78±1.642 ^{ab}
В	7.13±0.100 ^a	21.51±1.800a	3.17±0.289 ^b	1.08±0.090 ^{abc}	1.58±0.144 ^{bcd}	65.76±1505 ^b
С	7.20±0.200ª	22.09±2.461a	3.00±0.000 ^b	0.80±0.108 ^d	1.50±0.000 ^{cd}	65.47±2.829 ^b
D	7.20±0.200°	18.27±2.117 ^{ab}	3.67±0.289 ^a	1.01±0.045 ^{bcd}	2.00±0.000 ^d	67.86±2.305 ^{ab}
E	7.47±0.577°	19.87±3.778 ^{ab}	4.00±0.000 ^a	0.99±0.189 ^{bcd}	1.42±0.144 ^{cd}	67.52±4.216 ^{ab}
F	7.13±0.500 ^a	20.13±1.048 ^{ab}	2.67±0.289 ^d	0.91±0.106 ^{bcd}	1.33±0.144 ^d	67.40±0.980 ^{ab}
G	7.30±0.321ª	16.71±0.567 ^b	3.50±1.000 ^{ab}	0.92±0.134 ^{cd}	1.75±0.500 ^{abc}	69.81±2317 ^{ab}
Н	7.87±0.416 ^a	17.33±1.444 ^b	3.50±0.500a ^{bc}	0.87±0.072 ^{cd}	1.75±0.250 ^{abc}	69.42±2.005 ^{ab}
I	6.67±1.102ª	18.49±2.677 ^{ab}	2.83±0.289 ^{cd}	0.84±0.283 ^d	1.42±0.144 ^{abc}	68.56±2.580 ^{ab}
J	6.67±1.102 ^{ab}	16.09±2.155ª	2.83±0.289 ^{ab}	1.23±0.178 ^d	1.83±0.144 ^{cd}	71.35±3.421 ^b

Table 2. Proximate composition of fermented pigeon pea flour

Sample A, B and C: Pigeon pea fermented with Rhizopus nigricans at 24, 48 and 72 h, respectively Sample D, E and F: Pigeon pea fermented with Rhizopus oligosporus at 24, 48 and 72 h, respectively Sample G, H and I: Pigeon pea fermented with Rhizopus oryzae at 24, 48 and 72 h, respectively

Sample J: Control sample

4. CONCLUSION

Pigeon pea (*Cajanus cajan*) fermented using different *Rhizopus* species has been investigated for its functional and proximate properties. Improvement in functional and proximate properties of fermented pigeon pea flour were observed. Therefore, the product has good potential to serve as functional ingredient and supplement in different food formulations

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Mir SA, Shah MA, Ganai SA, Ahmad T, Gani M. Understanding the role of active components from plant sources in obesity management. Journal of the Saudi Society of Agricultural Sciences. 2017;16(11):25-32.
- 2. Venkidasamy B, Selvaraj D, Nile AS, Ramalingam S, Kai G, Nile SH. Indian pulses: A review on nutritional, functional and biochemical properties with future perspectives. Trends in Food Science and Technology. 2019;88:228-242.
- 3. Alfaro L, Chanda A, Kalemli-Ozcan SB, Sayek S. FDI and economic growth: the role of local financial markets. Journal of International Economics. 2004;(64)1:89-112.
- 4. Boukida F, Zannini E, Carini E, Vittadini E. Pulses for bread fortification: a necessity or a choice? Trends in Food Science and Technology. 2019;88:416-428.
- Feng C, Jin S, Luo M, Wang W, Xia XX, Zu YG, Fu YJ. Optimization of production parameters for preparation of natto-pigeon pea with immobilized Bacillus natto and sensory evaluations of the product. Innovative Food Science and Emerging Technology. 2015;31:160-169.
- Lee BH, Lai YS, Wu SC. Antioxidation, angiotensin converting enzyme inhibition activity, nattokinase, and antihypertension of *Bacillus subtilis* (natto)-fermented pigeon pea. Journal of Food and Drug Analysis. 2015;(23)4:750-757.
- 7. Olagunju AI, Omoba OS, Enujiugha VN, Alashi AM, Aluko RE. Pigeon pea enzymatic protein hydrolysates and ultrafiltration peptide fractions as potential

sources of antioxidant peptides: An in vitro study. LWT. 2018;97:269-278.

- Solomon SG, Okomoda VT, Oda SO. Nutritional value of toasted pigeon pea (*Cajanus cajan*) seed and its utilization in the diet of Clarias gariepinus (Burchell, 1822) fingerlings. Aquaculture Reports. 2017;7:34-39.
- Martínez-Villaluenga C, Torres A, Frias J, Vidal-Valverde C. Semolina supplementation with processed lupin and pigeon pea flours improve protein quality of pasta. LWT-Food Science and Technology. 2010;(43)4:617-622.
- Corriher VA, Hill GM, Bernard JK, Jenkins TC, West JW, Mullinix Jr BG. Pigeon peas as a supplement for lactating dairy cows fed corn silage-based diets. Journal of Dairy Science. 2010;(93)11:5309-5317.
- Srijunthongsiri S, Pradipasena P, Tulyathan V. Influence of heat-moisture modification in the presence of calcium compound on physicochemical properties of pigeon pea [*Cajanus cajan (L.)* Millsp.] starch. Food Hydrocolloids. 2016;53:192-198.
- 12. Yadav BS, Yadav RB, Kumar M. Suitability of pigeon pea and rice starches and their blends for noodle making. LWT-Food Science and Technology. 2011;(44)6: 1415-1421.
- Anuonye JC, Ndaliman M, Elizabeth OU and Yakubu MC. Effect of blending on the composition and acceptability of blends of unripe banana and pigeon pea flours. Nigerian Food Journal. 2012;(30)1:116-123.
- Balasubramanian A, Durairajpandian V, Elumalai S, Mathivanan N, Munirajan AK, Ponnuraj K. Structural and functional studies on urease from pigeon pea (*Cajanus cajan*). International Journal of Biological Macromolecules. 2013;58:301-309.
- Qi XL, Li TT, Wei ZF, Guo N, Luo M, Wang W, Peng X. Solvent-free microwave extraction of essential oil from pigeon pea leaves [*Cajanus cajan (L.)* Millsp.] and evaluation of its antimicrobial activity. Industrial Crops and Products. 2014;58: 322-328.
- Olanipekun BF, Otunola ET, Oyelade OJ. Effect of fermentation on antinutritional factors and in vitro protein digestibility of bambara nut (*Voandzeia subterranean L.*) Journal of Food Science and Quality Management. 2015;39:98- 110.

- Owuamanam CI, Ogueke CC, Achinewhu SC, Barimalaa IS. Quality characteristics of gari as affected by preferment liquor, temperature and duration of fermentation. American Journal of Food Technolonogy. 2011;6(5):374-384.
- Nwanekezi EC, Ohagi NC, Oc AA. Nutritional and organoleptic quality of infant food formulations made from natural and solid state fermented tubers (cassava, sprouted and unsprouted yam)-soybean flours blend. Nigerian Food Journal. 2001; 19:55-62.
- 19. Tester RF, Morrison WR. Swelling and gelatinization of cereal starches. I. Effects of amylopectin, amylose and lipids. Cereal Chemistry. 1990;67:551.
- 20. AOAC. Official methods of analysis, association of official analytical chemists.17th ed. Washington DC; 2005.
- Adelakun OE, Olanipekun BF, Akingbaso O, Adhikar BM. Effect of fermentation and variety on quality attributes of Okra seed (*Abelmoshus esculentus L*. Moench) flour. Donnish Journal of Food Science and Technology. 2017;3(1):001-006.
- Olanipekun OT, Obatolu VA, Ejoh SI, Oyarekua MA, Farinde EC. Evaluation of the nutrients and antinutrient contents of two Nigerian under-utilized legumes. Moor Journal of Agricultural Research. 2011;12: 56 - 63.
- 23. Oyelade OJ, Ade-Omowaye BIO, Adeomi VF. Influence of variety on protein, fat contents and some physical characteristics of okra seeds. Journal of Food Engineering. 2003;57(2):111-114.
- Lawal OS, Afolabi TA, Adebowale KO, Ogunsanwo BM, Bankole SA. Effect of selected Hofmeister anions on functional properties of protein isolate prepared from lablab seeds (*Lablab purpureus*). Journal of the Science of Food and Agriculture. 2005;85(15):2655-2659.
- 25. Adebowale OJ, Maliki K. Effect of fermentation period on the chemical composition and functional properties of Pigeon pea (*Cajanus cajan*) seed flour. International Food Research Journal. 2011;18(4):1329 1333.
- Edema MO, Sanni LA, Sanni AI. Evaluation of maize-soybean flour blends for sour maize bread production in Nigeria. African Journal of Biotechnology. 2005;4: 911–918.
- 27. Onimawo IA, Momoh AH. Usman A. Proximate composition and functional

properties of four cultivars of bambara groundnut. Plant Foods for Human Nutrition. 1998;53(2):153-158.

- Oshodi HN, Ogungbenle MO, Oladimeji AA. Chemical composition, nutritionally valuable minerals and functional properties of benniseed (*Sesamum radiatum*), pearl millet (*Pennisetum typhoides*) and quinoa (*Chenopodium quinoa*) flours. International Journal of Food Sciences and Nutrition. 1999;50(5):325-331.
- 29. [Fasasi OS. Proximate anti-nutritional factors and functional properties of processed pearl millet (*Pennisetum glaucum*). Journal of Food Technology. 2009;7(3):92-97.
- Sefa-deheh S, Sanni AI, Tetteh G, Sakyi-Dawson E. Yeasts in the traditional brewing of *pito* in Ghana. World Journal of Microbiology and Biotechnology. 1999; 15:593–597.
- Emmanuel I, Aladesanmi O, Adekunle O, Oluwafemi O. Effect of Fermentation on the Physicochemical Properties and Nutritionally Valuable Minerals of Locust Bean (*Parkia biglobosa*), American Journal of Food Technology. 2017;12(6):379-384.
- 32. Lucretia IB, Patience CO, Confidence IE. Proximate Composition and Sensory Properties of Complementary Food Formulated from Malted Pre-Gelatinized Maize, Soybean and Carrot Flours. Journal of Food Research. 2018;7(2):17-26.
- Adebowale AA, Sanni LO, Owo HO, Karimm OR. Effect of variety and moisture content on some engineering properties of paddy rice. Journal of Food Science and Technology. 2011;48(5):551-559.
- 34. Anthony O, Babatunde B. Effect of Fermentation on Nutrient and Antinutrients composition of millet (*Pennisetum glaucum*) and soya bean (*Glycine max*) blend of flours. Journal of Life Sciences. 2014;8(8):668-675.
- Ahmed Z, Wang Y, Xu N, Xi A, Zang B, Bai X. Effects of Lactobacillus plantarum MA2 isolated from Tibet kefir on lipid metabolism and intestinal microflora of rats fed on high-cholesterol diet. Journal of Applied Microbiology and Biotechnology. 2009;84:341-7.
- 36. Ojokoh AO, Abiola AB, Lawal RT. Changes in nutrient and antinutrient composition of popcorn and groundnut composite flour subjected to solid substrate fermentation. African Journal of

Agricultural Research. 2012;7(23):3439-3445.

37. Omafuvbe BO, Falade OS, Osuntogun BA, Adewusi SRA. Chemical and biochemical changes in African locust bean (Parkia biglobosa) and Melon (Citrullus vulgaris) Seeds during Fermentation to Condiments. Pakistan Journal of Nutrition. 2004;3(3): 140-145.

 Onweluzo JC, Nnamuchi OM. Production and evaluation of porridge-type breakfast product from Treculia africana and sorghum bicolor flours. Pakistan Journal of Nutrition. 2009;8(6):731-736.

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